

# Nitrous Oxide Production in an Eastern Corn Belt Soil: Sources and Redox Range



G. Hernandez R., S.M. Brouder, D.R. Smith, G.E. Van Scoyoc, T.R. Filley, and G. Michalski

915 W State St, West Lafayette, IN / E-mail: hermandg@purdue.edu



## Justification

- Nitrous oxide ( $N_2O$ ) is a main greenhouse gas and destroyer of ozone layer.
- Sources and regulation of  $N_2O$  production are not well understood.
- Agroecosystems are an important, highly-dynamic source of  $N_2O$ .
- Discrimination of soil  $N_2O$  sources may enhance our prediction ability of regional and global  $N_2O$  emissions.
- The interactive effect among increasing soil water content, redox potential (Eh), agricultural N management practices and crop growth stage on  $N_2O$  production need to be examined in detail.
- New knowledge about  $N_2O$  production may enhance N management and efficiency for different N sources (e.g.: manure vs. synthetic).



Experimental site (WQFS - Purdue University): corn plot.

## Objective

- To estimate magnitude and sources of  $N_2O$  production as affected by N management, soil water content, Eh and corn growth stage.

## Hypothesis

- Urea-ammonium nitrate (UAN) favors nitrification and liquid swine manure (M) favors denitrification.
- Manured soils have a higher, narrower Eh range of  $N_2O$  production than soils receiving UAN.

## Materials and Methods

### Experimental Site and Treatment Management:

- The Water Quality Field Station (WQFS) at Purdue University – Agronomy Center for Research and Education (ACRE) on Drummer (Typic Endoaquoll) and Raub (Aquic Argiudoll) soil series.
- Continuous corn (CC) fertilized with UAN (28% N)  $157 \text{ kg N ha}^{-1}$  at V5, or liquid swine manure (M) at  $255 \text{ kg N ha}^{-1}$  in the fall (F). Treatments were CCFM and CCUAN in a randomized complete block design.

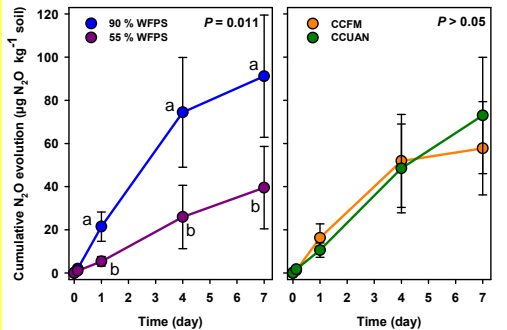
### Soil Sample Collections and Incubation Experiments:

- Soil cores (n: 18) were collected (0 – 15 cm depth) at preplant (PP) and at V6, sieved to aggregate size ( $< 6.4 \text{ mm}$ ), packed to  $1.2 \text{ g cm}^{-3}$ , and preincubated (3 d) at 45 % water filled pore space (WFPS).
- I. Aerobic Incubation (7 d): 90 g soil (oven-dried equivalent, OVDE) in 0.97 L Mason jars (n: 96)
- II. Aerobic Incubation – Source-partitioning (24 hr): 90 g soil (OVDE) in 0.97 L Mason jars (n: 64)
  - Labeling (20 atom excess a.e.%) N pools with  $^{15}\text{N-NH}_4^+$  for nitrification and  $^{15}\text{N-NO}_3^-$  for denitrification
  - Primary assumption: uniform pool labeling within each microcosm for both ions.
  - Assuming natural abundance: 0.3663 % ( $^{15}\text{N}$  a.e.). Sources expressed as % of  $N_2O$  production at 24 h.
- III. Anaerobic Incubation (15 d): 60 g soil (OVDE) in 0.25 L centrifuge bottles (n: 12). Degassed  $\text{H}_2\text{O}$ :soil 2.3:1. Amendment:  $\text{KNO}_3$  at  $50 \text{ mg N kg}^{-1}$  soil. Pure  $\text{N}_2$  flow in headspace. Electrode: Ag/AgCl sat. KCl.
  - Eh values were corrected to standard  $\text{H}_2$  electrode (+ 199 mV) and to pH 7.
- Experimental factors in the incubation were soil moisture (55 and 90 % WFPS or submerged), 2 trt (CCFM and CCUAN), 2 soil sample collections (PP and V6) and 4 field replicates (blocks)

### Analysis Procedures:

- Extractable dissolved organic carbon (EDOC): 20 g (OVDE) of air-dried soil in 100 mL (5 mM  $\text{CaCl}_2$ ), shaking, centrifugation, filtration and TOC analyzer. Soil organic C (SOC) by dry combustion.
- Nitrous oxide analysis: GC with ECD. ConFlow III GC/IRMS for  $\delta^{15}\text{N}$  in  $\text{N}_2\text{O}$  with a cryotrap.
- Statistics: Cook's distance for outliers, VIF for multicollinearity, adjusted  $R^2$  for variable selection, Linear Regression, RM ANOVA and Tukey test for pairwise comparisons.

## Results Part I: Soil Moisture and N Source Impacts on N<sub>2</sub>O Production (Aerobic Incubation)

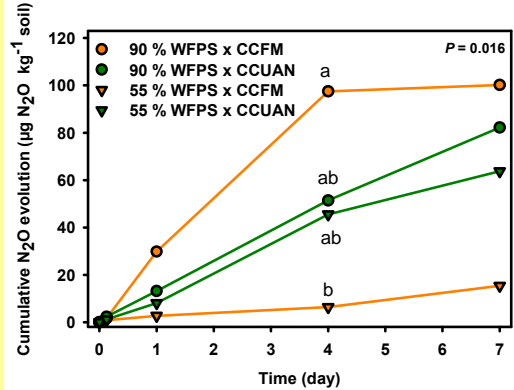


### Individual Factor Effects on N<sub>2</sub>O Production:

- Much greater N<sub>2</sub>O production (2.3 times) with high WFPS due to limited O<sub>2</sub> availability.
- N management effect was not significant.
- Greater CO<sub>2</sub> production (1.3 times at 7<sup>th</sup> d) with 90 than 55 % WFPS ( $P < 0.01$ , data not shown).

### Interaction Effect on N<sub>2</sub>O Production:

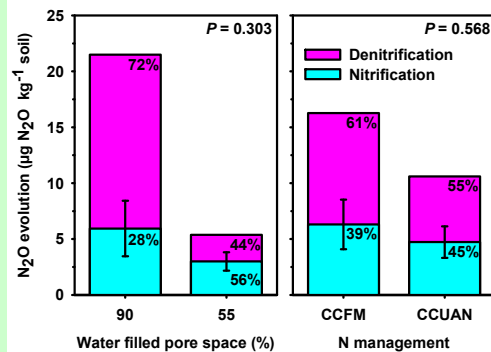
- Among the 4 treatment combinations, the highest, shortest-lived N<sub>2</sub>O production occurred with manured soils at high water content.



## Results Part II: N<sub>2</sub>O Source-partitioning (Aerobic Incubation)

### N<sub>2</sub>O Derived from Nitrification vs. Denitrification:

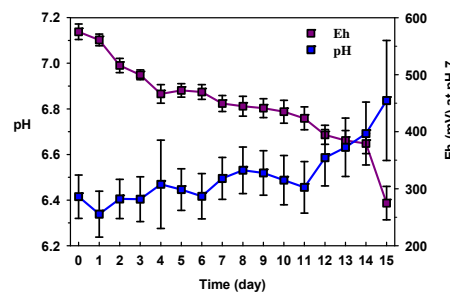
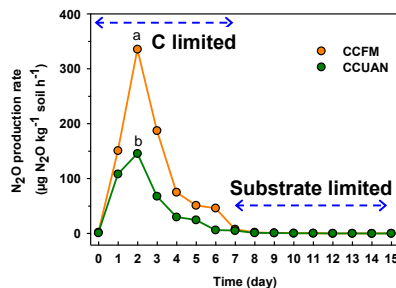
- Non-significant results for both factors, but clear tendency of greater denitrification proportion (from 44 to 72 %) with increasing soil water content.
- SOC and EDOC were 1.07 ( $P = 0.023$ ) and 1.21 ( $P = 0.046$ ) times greater in CCFM than CCUAN (data not shown). Higher EDOC in CCFM also supports denitrification as main pathway for N<sub>2</sub>O production.
- Also, greater CO<sub>2</sub> production (1.2 times at 7<sup>th</sup> d) in CCFM than in CCUAN ( $P = 0.057$ , data not shown).
- Microsite anaerobiosis favored by increased microbial respiratory demand of O<sub>2</sub> in CCFM may have enhanced denitrification.



## Results Part III: Redox Potential Impacts on N<sub>2</sub>O Production (Anaerobic Incubation)

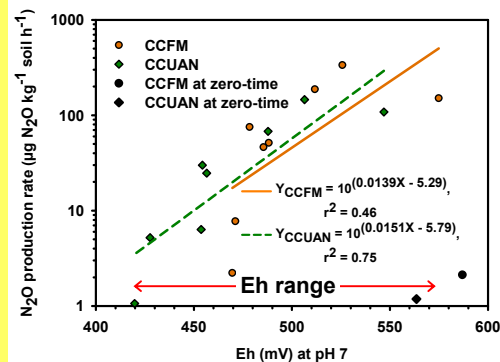
### N<sub>2</sub>O Production Rates, Eh and pH Patterns with Time:

- N<sub>2</sub>O production rate was 2.3 times higher in CCFM than in CCUAN on day 2 ( $P = 0.018$ ).
- Most N<sub>2</sub>O was produced within the first four days of incubation coinciding with a sharp drop in Eh.
- As expected, when Eh progressively declined with time, pH tended to increase ( $r: -0.89$ ,  $P < 0.001$ ).



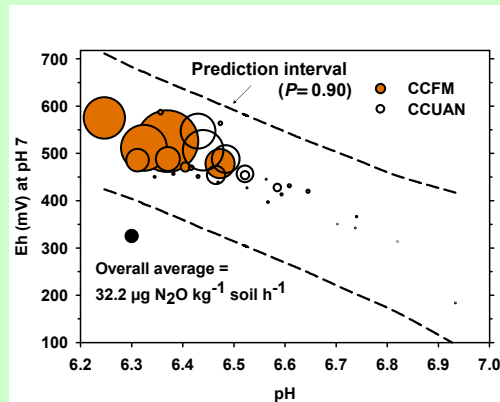
### Redox Potential Range for N<sub>2</sub>O Production:

- N fertilizer source caused minimal treatment differences between N<sub>2</sub>O production rate patterns:
  - In general, Eh range for N<sub>2</sub>O production was not affected by N management.
  - Although fall manured soils (470 to 575 mV) tended to have a range shifted to higher Eh values than soils receiving UAN (420 to 550 mV), the two Eh ranges overlapped each other.
- The two N fertilizer treatments showed similar, sharp exponential declining phase of N<sub>2</sub>O production rates with decreasing Eh.



### Eh Prediction Interval with Varying pH:

- Bubble areas correspond to N<sub>2</sub>O production rates.
- Higher N<sub>2</sub>O production was at high Eh and low pH.
- Linear regressions:
  - Eh = 3400 - 453pH ( $r^2 = 0.78$ ,  $P < 0.001$ ).
  - N<sub>2</sub>O production rates = 3.18 - 0.47pH ( $P < 0.01$ ).
  - N<sub>2</sub>O production rates = -0.309 + 0.000936Eh ( $P < 0.01$ ).
- Redundancy between pH and Eh as predictors was demonstrated by moderate collinearity (VIF > 4.5).
- The interactive, pH-Eh-N<sub>2</sub>O pattern suggested enhanced prediction ability when combining the two master variables (pH and Eh).
- N fertilizer did not impact the pH-Eh-N<sub>2</sub>O pattern.



## Conclusions

- Increasing soil water content and fall liquid swine manure showed a synergistic interaction that enhances soil N<sub>2</sub>O production in aerobic conditions.
- Extreme increases of soil water content favored denitrification as predominant pathway of the largest pulse of N<sub>2</sub>O production in aerobic conditions.
- In anaerobic conditions, fall manured soils registered greater N<sub>2</sub>O production than soils receiving side-dressed urea-ammonium nitrate shortly after flooding.
- The first 4-days of our incubations showed the highest magnitude and dynamic of N<sub>2</sub>O production.
- Redox potential range (420 to 475 mV) for soil N<sub>2</sub>O production was not affected by N fertilizer source.
- No temporal effects (corn preplanting vs. growth stage V5) were observed in our experiments.
- Additional variable screening for new covariates (e.g. soil microbial biomass C, light POM C, electrical conductivity), model development and data validation are necessary for better understanding of N<sub>2</sub>O production.
- Future work may include the impact of soil aggregate size, microsite diversity, entrapped air and/or root exudates on magnitude and sources/sinks of soil N<sub>2</sub>O production/consumption.
- Headspace acetylene additions and isotopomers (positional preference) techniques may also be helpful in N<sub>2</sub>O source-partitioning studies.
- Assessing dissolved N<sub>2</sub>O production in anaerobic incubations may enhance our current understanding on N<sub>2</sub>O emissions from flooded soils.

## Acknowledgements:

Funding by – Soil and Soil Biology Program (USDA NRI)  
– Consortium for Agricultural Soils Mitigation of Greenhouse Gases (CASMGs)



Thanks to lab technicians and helpers:  
Amanda, Lucas, Jennifer, Jeff, Rhonda,  
Dennis, Brenda and Marianne