

Identification and Characterization of Flowering Time Genes in Tall Wheatgrass

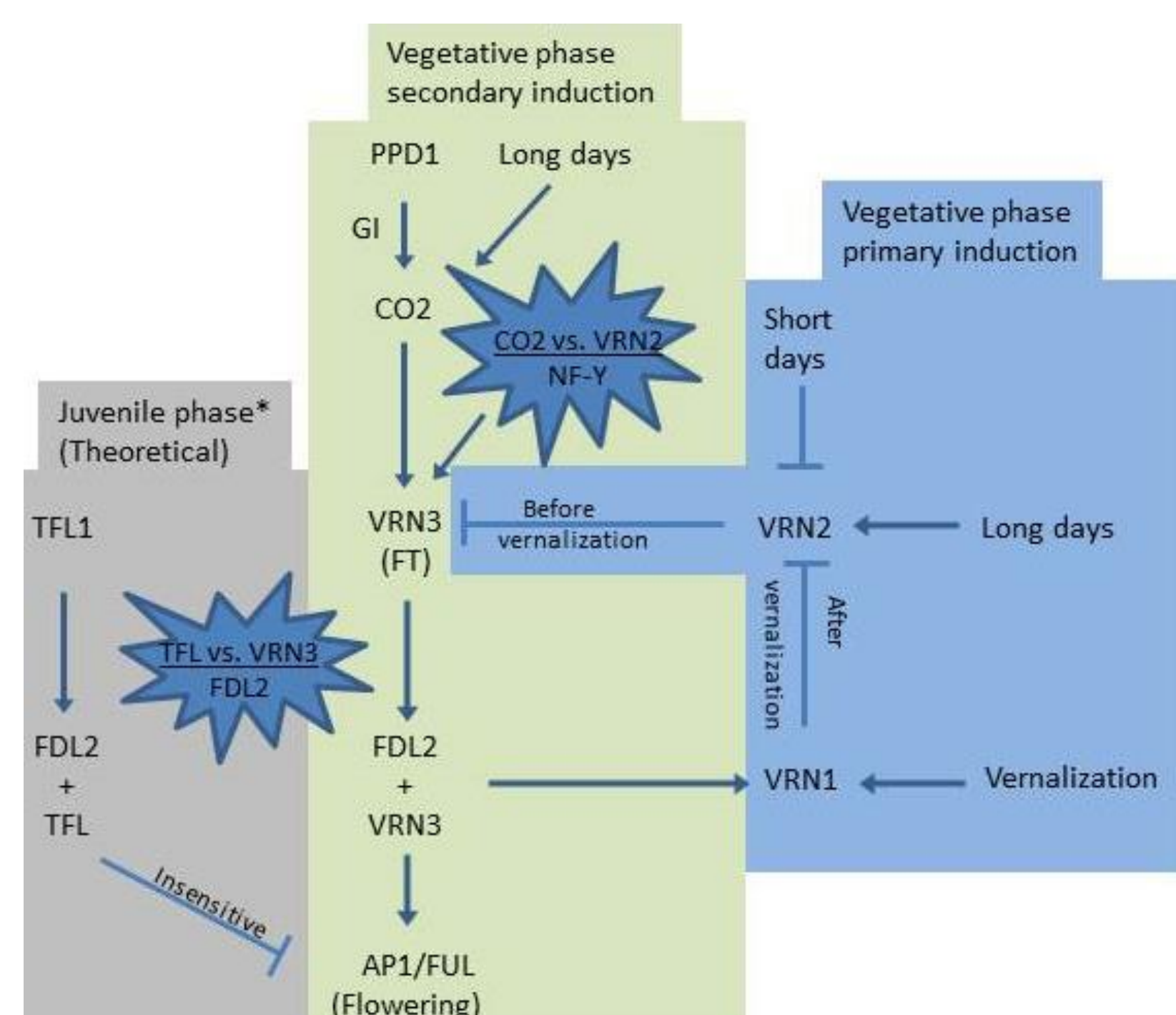
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Introduction

Compared to annuals, perennial cereals confer significant environmental and economic benefits [1]. Tall wheatgrass, *Thinopyrum elongatum* and *Th. ponticum*, are long lived perennial grasses closely related to wheat and provide an important source of perennial habit for the development of perennial wheat. Despite the large amount of molecular information in annuals, limited plant development and molecular information is available for *Thinopyrum sp.* With the existing knowledge of flowering time pathways in annual and perennial plant species, we identified key flowering time candidate genes for sequencing and expression analysis.



Candidate genes	Expressed in
<i>VRN1</i> (<i>VERNALIZATION1</i>)	Leaf and meristems
<i>VRN2</i> (<i>VERNALIZATION2</i>)	Leaf
<i>VRN3</i> (<i>VERNALIZATION3</i>)	Leaf and meristems
<i>CO2</i> (<i>CONSTANS2</i>)	Leaf
<i>PPD1</i> (<i>PHOTOPERIOD1</i>)	Leaf
<i>FDL2</i> (<i>FD LIKE2</i>)	Leaf and meristems
<i>TFL1</i> (<i>TERMINAL FLOWER1</i>)	meristems
<i>GI</i> (<i>GIGANTEA</i>)	Leaf

Vegetative to reproductive phase transition requires a co-ordinated action of vernalization and photoperiod. While *VRN1*, a MADS-box meristem identity gene, is up-regulated by vernalization [2], *VRN2*, the central repressor of flowering is down-regulated [3]. *VRN3* (TaFT) is induced by long days once vernalization requirement is met [4]. As such, *CO2*, which is up-regulated in long days, outcompetes *VRN2* to create a complex with NF-Y proteins and this *CO2*-NF-Y complex promotes *VRN3* expression [5]. Further, *GI*, which encodes a nuclear-localized membrane protein, functions upstream of *CO2* and *FT* [6]. *PPD1*, activated by phytochrome C, is required for high levels of *FT* (*VRN3*) under long days [7] which in turn induces *VRN1* expression, through interaction with *FDL2* [8], creating a positive feedback loop. In *Arabidopsis*, *TFL1* is antagonistic to *FT* and is proposed to compete for binding with *FDL2* to repress flowering. Reduced *TFL1* expression in *Arabis alpina* (perennial weed) is linked to shorter juvenile period [9] suggesting its potential role in perennial habit.

Results and discussion

Genes	Expected length for Chinese Spring	<i>Thinopyrum</i> Accessions	Length of consensus sequence	% similarity to Chinese spring	Major insertion/deletions w.r.t. Chinese spring	Cladogram	Progress		
							Gene	Coding region	5' upstream region
<i>CO2</i>	647	PI 531717	647	99.5	Highly conserved.		<i>VRN1</i>	Sequenced	In process
		PI 531718	647	99.5					
		PI 531719	647	99.3					
		PI 206624	647	99.5					
		Chinese spring	647	99.3					
<i>VRN2</i>	592	PI 531717	613	84.5	Insertion of bases in multiples of 3 without changes to the conserved CCT domain.		<i>VRN2</i>	Sequenced	In process
		PI 531718	613	83.8					
		PI 531719	604	82.5					
		PI 206624	616	92.9					
		Chinese spring	613	84.5					
<i>FDL2</i>	375	PI 531717	375	98.4	Conserved with few SNPs.		<i>VRN3</i>	In process	In process
		PI 531718	375	98.8					
		PI 531719	375	96.5					
		PI 206624	375	97.5					
		Chinese spring	375	98.4					
<i>VRN1</i>	701	PI 531717	701	97.8	Conserved with few SNPs.		<i>PPd1</i>	Sequenced	In process
		PI 531718	701	97.8					
		PI 531719	701	97.2					
		PI 206624	701	97.6					
		Chinese spring	701	97.8					
<i>Ppd1</i>	1918	PI 531717	2068	87.8	Insertion or deletion of bases in multiples of 3 without changes to the conserved Response-regulatory and CCT domains.		<i>CO2</i>	Sequenced	In process
		PI 531718	2062	87.9					
		PI 531719	1879	93.0					
		PI 206624	1882	92.4					
		Chinese spring	1918	87.8					
<i>TFL1</i>	429	PI 531717	429	97.2	Conserved with few SNPs.		<i>FDL2</i>	Sequenced	Sequenced
		PI 531718	429	97.8					
		PI 531719	429	97.7					
		PI 206624	429	96.9					
		Chinese spring	429	97.2					

A 144 bp insertion in PI 531717 and PI 531718 for *Ppd1* gene aligns to a region of *Ppd-B1* exon 7 in *Triticum turgidum*, *T. durum*, etc.

Sequencing was completed at McGill University and the Genome Quebec Innovation centre. Sequence alignments, computation of similarity and cladogram constructions were all performed in *geneious 6.1.6* software.

References

[1] Bell et al (2010). Crop Pasture Sci. 61: 679-690. [2] Yan L. et al (2003). PNAS 100(10): 6263-68. [3] Yan L. et al (2004). Science 303(5664): 1640-44. [4] Yan L. et al (2006). PNAS 103(51): 19581-86. [5] Li C et al (2011). Plant J. (67): 763-73. [6] Tseng TS et al (2004). Plant cell 16(6): 1550-63. [7] Chen A et al (2014). PNAS 111(28): 10037-44. [8] Li C and Dubcovsky J (2008). Plant J 55(4): 543-54. [9] Wang R et al (2011). Plant Cell 23(4):1307-21.

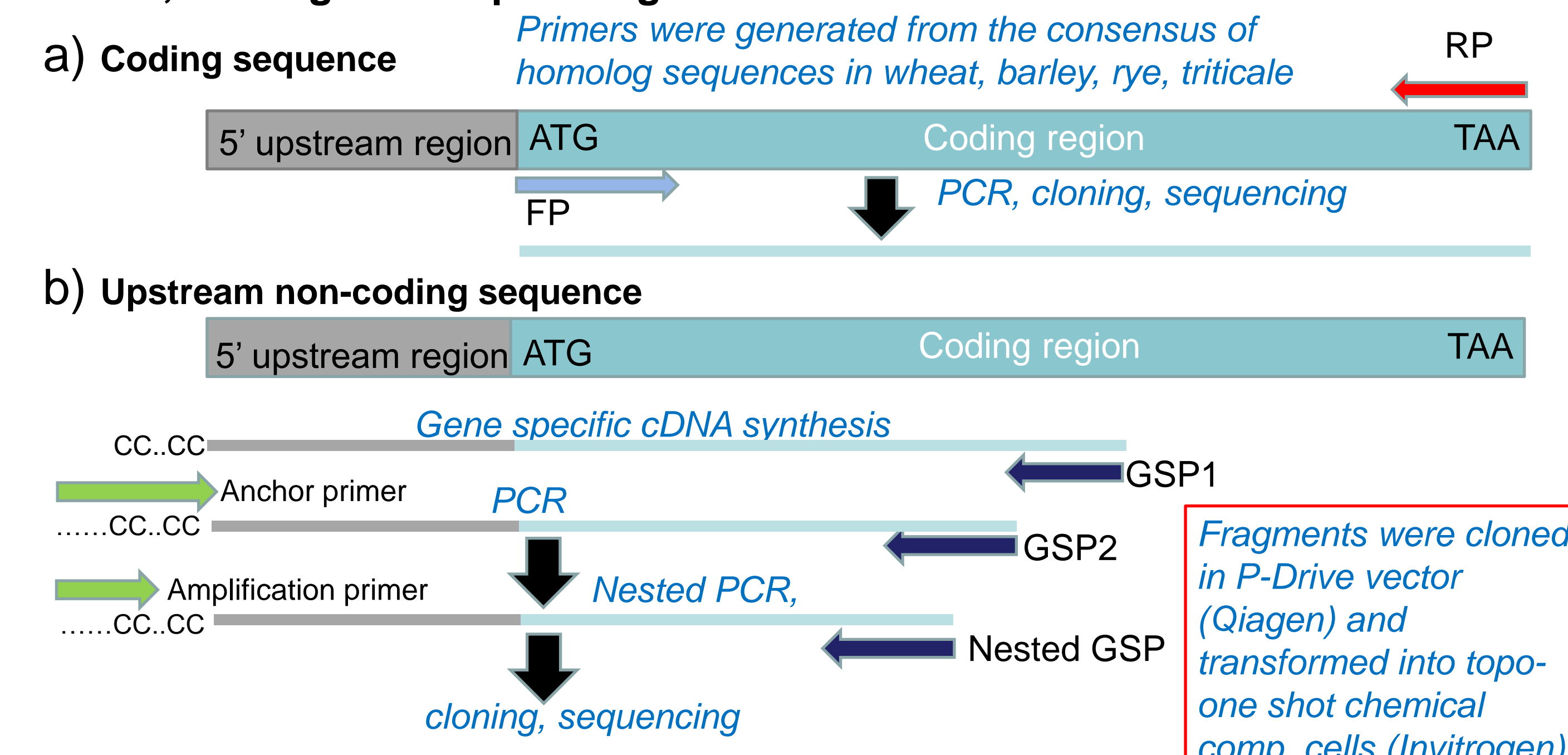
Material and methods

1. Plant materials:

Species	Accessions	Ploidy level	Growth habit
<i>Triticum aestivum</i>	Chinese spring	2n=6X=42	Spring, annual
<i>Thinopyrum elongatum</i>	PI 531717	2n=2X=14	Spring, perpetual flowering
	PI 531718	2n=2X=14	Spring, perpetual flowering
	PI 531719	2n=8X=56	Winter, perennial
<i>Thinopyrum ponticum</i>	PI 206624	2n=10X=70	Winter, perennial

2. RNA extractions: Total RNA was extracted from young leaves and/or meristems (3 leaf stage)

3. PCR, cloning and sequencing



4. Next steps

Genome walking will be performed to elucidate the genetic makeup of the intron regions. The expression data and effects of the candidate genes on phenotype and flowering will be carried out through quantitative RT-PCR and virus induced gene silencing (VIGS) respectively.