

# Multiple untranslated exons and introns might be related to differences of expression in barley S-adenosylmethionine synthetase genes

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## Absrtact

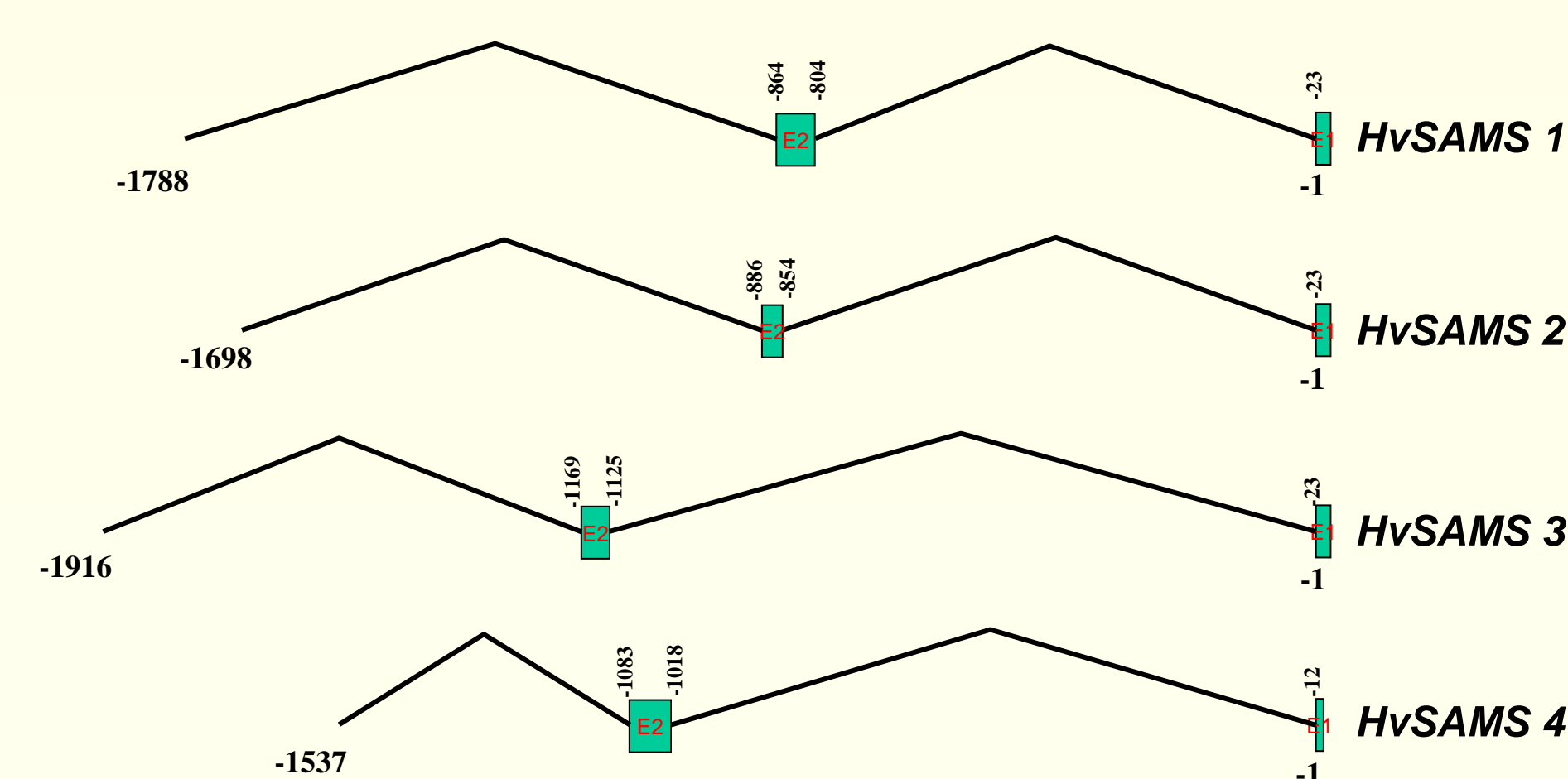
Barley S-adenosylmethionine synthetase (*HvSAMS*) catalyses the biosynthesis of S-adenosylmethionine (SAM) from ATP and L-methionine. *HvSAMS 1, 2, 3, and 4* genes and their 5' upstream regions were isolated and analyzed. The genomic DNA sequences of *HvSAMS 1, 2, 3, and 4* genes did not contain intron. However, two introns and two exons were present on the 5' upstream regions of each *HvSAMS 1, 2, 3, and 4*. They were present within 1787 bp (*HvSAMS 1*), 1698 bp (*HvSAMS 2*), 1916 bp (*HvSAMS 3*), and 1537 bp (*HvSAMS 4*) from the translated initiation site.

Although high sequence homology was found among the four *HvSAMS* genes and first exon on the 5' upstream region, numerous deletions and insertions were found in the second exon from the 5' upstream region and two introns. The second exon from the 5' upstream region among *HvSAMS 1, 2, 3, and 4* showed high structural variation. In order to identify proposed alternative splicing occurred in 5' upstream region, RT-PCR with exons and introns of promoter specific primers were conducted using barley grains of different developmental stages. Presence of different transcript sequences derived from the 5' upstream region might indicate different upstream sequence use of *HvSAMS* genes.

## METHODS

- Plant material : early maturity barley germplasm "GSHO 2504" *Arabidopsis thaliana* (Col-0)
- Promoter isolation : Marathon cDNA amplification kit (Clontech) Universal GenomeWalker (Clontech)
- Transient expression assay : deletion vector series (Fig. 3) *Agrobacterium tumefaciens* (GV3101)
- Analysis : Blast alignment, PLACE web signal scan, and PlantCARE RT-PCR  
GUS activity assay  
Histochemical staining

## RESULTS



**Fig. 1** Schematic representation of the 5' UTRs of the *HvSAMS 1, 2, 3, and 4*. Solid bars represented exons (E1 and E2), whereas introns were indicated by the straight/"A" lines. The nucleotide of the ATG translation initiation codon was assigned as position 1 in the nucleotide sequence, and the nucleotide positions upstream of position 1 were presented as negative numbers.

**Table 1** Analysis of the *HvSAMS 1, 2, 3, and 4* of 5' untranslated regions. The PlantCare was used for each *cis*-elements.

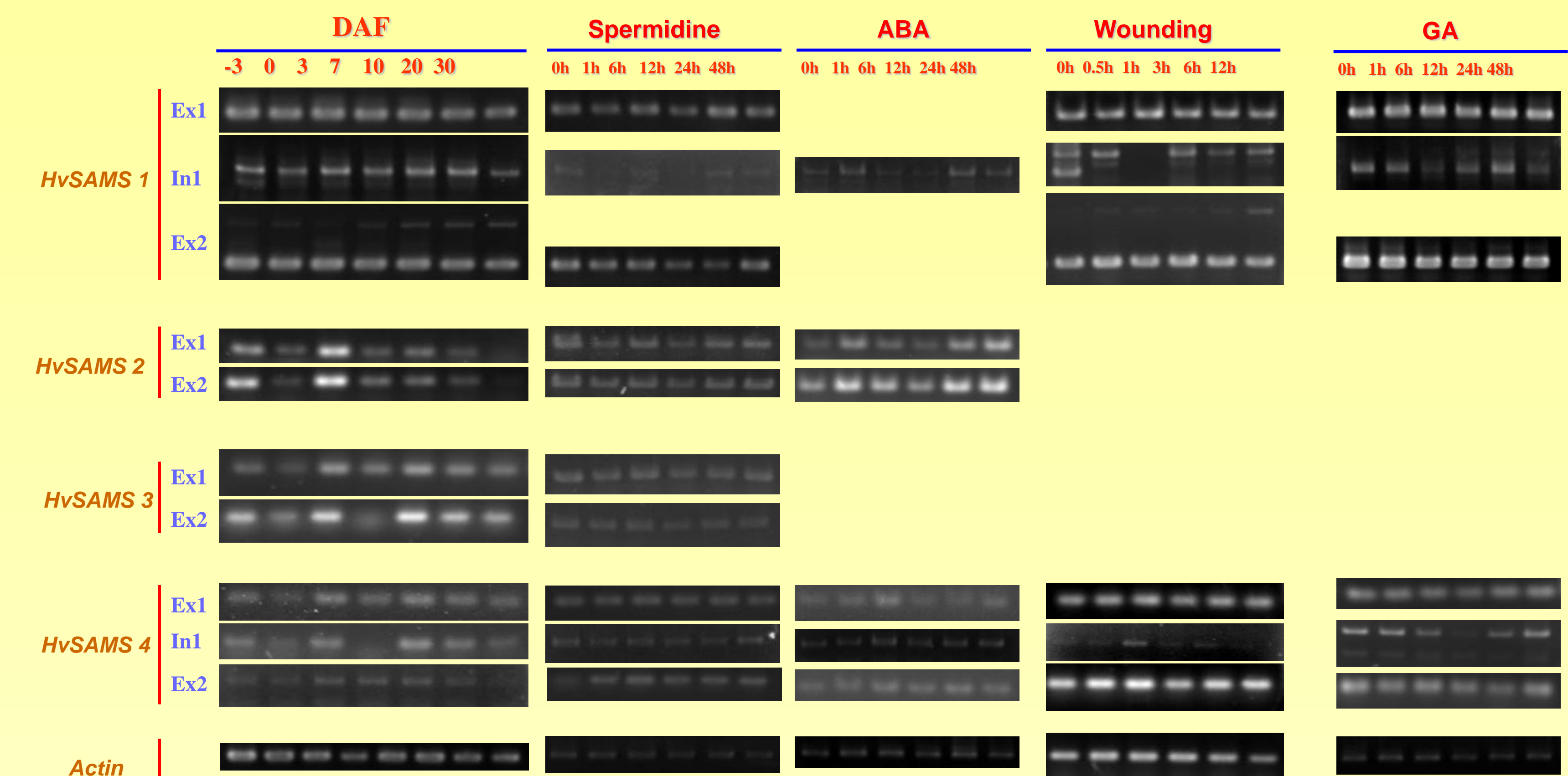
Gene	cDNA <sup>a</sup>		genomic DNA <sup>b</sup>		Conserved sequence	Function
	cis-element	Location	cis-element	Location		
<i>HvSAMS 1</i>	AAGAA-motif	(+178)	AAGAA-motif	(-1601 to +1528) (-1553)	GAAAGAA	
	CAAT-box	(+137)	CAAT-box	(-920 to +1176) (-1176)	CAAT	common cis-acting element in promoter and enhancer regions
	EIRE	(+5)	EIRE	(-920 to +959)	TTGACC	elicitor-responsive element
	LTR	(-114)	LTR	(-968 to +480)	CGAAA	cis-acting element involved in low-temperature responsiveness
	MBS	(+51)	MBS	(-1050 to +1050)	CAACTG	MYB binding site involved in drought-inducibility
	SARE	(+5)	SARE	(-959 to +1562)	TTGACCTCCTT	cis-acting element involved in salicylic acid responsiveness
	circadian	(+137)	A-box	(-1515 to +1367)	GAANNATC	cis-acting regulatory element involved in circadian control
			C-box	(-1515 to +1367)	CTGACGTCAG	cis-acting regulatory element involved in light responsiveness
			G-box	(-891 to +675) (-1756)	CACGTT	cis-acting regulatory element involved in light responsiveness
			TCA-element	(+453 to +1519)	CCATCTTTT	cis-acting element involved in salicylic acid responsiveness
<i>HvSAMS 2</i>			TGACG-motif	(+1588 to +1581)	TGACG	cis-acting regulatory element involved in the MeJA-responsiveness
	CAAT-box	(+9)	CAAT-box	(-1064 to +1484) (-1289)	CAAT	common cis-acting element in promoter and enhancer regions
	GAG-motif	(-137)	MBS	(-845 to +983)	AGAGATG	light responsive element
	MBS	(+123)	MBS	(-845 to +983)	CAACTG	MYB binding site involved in drought-inducibility
	circadian	(+9)	circadian	(-831 to +1311)	CAANNATC	cis-acting regulatory element involved in circadian control
			A-box	(-1152 to +1152)	CGTCC	cis-acting regulatory element
			ABRE	(-988 to +1179)	TACGTG	cis-acting element involved in the abscisic acid responsiveness
			CGTCA-motif	(-1472 to +1472)	CGTCA	cis-acting regulatory element involved in the MeJA-responsiveness
			G-box	(-1168 to +1168)	CACGTA	cis-acting regulatory element involved in light responsiveness
			GCNA <sub>1</sub> -motif	(-628 to +308)	TGAGTCA	cis-regulatory element involved in endosperm expression
<i>HvSAMS 3</i>			LTR	(-1448 to +1481)	CGAAA	cis-acting element involved in low-temperature responsiveness
			Skn-1 <sub>1</sub> -motif	(-461 to +1360)	GTCA	cis-acting regulatory element required for endosperm expression
			TGA-element	(-1179 to +1472)	AACGAC	auxin-responsive element
			TGACG-motif	(+1584 to +1884)	TGACG	cis-acting regulatory element involved in the MeJA-responsiveness
	CAAT-box	(+29)	CAAT-box	(-1781 to +1147) (-1453)	CAAT	common cis-acting element in promoter and enhancer regions
	circadian	(+43)		(-876 to +1139)	CAANNATC	cis-acting regulatory element involved in circadian control
			ABRE	(-1139 to +1852)	TACGTG	cis-acting element involved in the abscisic acid responsiveness
			AuxRR-core	(-1854 to +1470)	GGTCCAT	cis-acting regulatory element involved in auxin responsiveness
			CGTCA-motif	(-1470 to +1141)	CGTCA	cis-acting regulatory element involved in the MeJA-responsiveness
			G-box	(-1491 to +1491)	CACGTA	cis-acting regulatory element involved in light responsiveness
<i>HvSAMS 4</i>			MBS	(-982 to +1587)	CGGTCA	MYB Binding Site
			Skn-1 <sub>1</sub> -motif	(-461 to +1360)	GTCA	cis-acting regulatory element required for endosperm expression
			TCA-element	(-1448 to +1481)	GAGAAGAATA	cis-acting element involved in salicylic acid responsiveness
			TGACG-motif	(-1364 to +1364)	TGACG	cis-acting regulatory element involved in the MeJA-responsiveness
			CGTCA-motif	(+123 to +1364)	CGTCA	cis-acting regulatory element involved in the MeJA-responsiveness
			Skn-1 <sub>1</sub> -motif	(-1364 to +1364)	GTCA	cis-acting regulatory element required for endosperm expression
			circadian	(-1054 to +1220)	CAANNATC	cis-acting regulatory element involved in circadian control
			ABRE	(-920 to +1305)	TACGTG	cis-acting element involved in the abscisic acid responsiveness
			AuxRR-core	(-951 to +1216)	GGTCCAT	cis-acting regulatory element involved in auxin responsiveness
			CAAT-box	(-987 to +1288)	CAAT	common cis-acting element in promoter and enhancer regions
<i>HvSAMS 4</i>			ERE	(-1390 to +1465)	ATTTCAAA	ethylene-responsive element
			G-Box	(-920 to +1538) (-1270)	CACGTA	cis-acting regulatory element involved in light responsiveness
			GCNA <sub>1</sub> -motif	(-628 to +157)	TGAGTCA	cis-regulatory element involved in endosperm expression
			LTR	(-1157 to +1157)	CGAAA	cis-acting element involved in low-temperature responsiveness
			MBS	(-1384 to +1384)	CAACTG	MYB binding site involved in drought-inducibility
			TCA-element	(+1162 to +1500)	GAGAAGAATA	cis-acting element involved in salicylic acid responsiveness
			TGA-element	(+297 to +1316)	AACGAC	auxin-responsive element
			TGACG-motif	(-1316 to +1316)	TGACG	cis-acting regulatory element involved in the MeJA-responsiveness

<sup>a</sup> The column was isolated from 5' RACE clones.

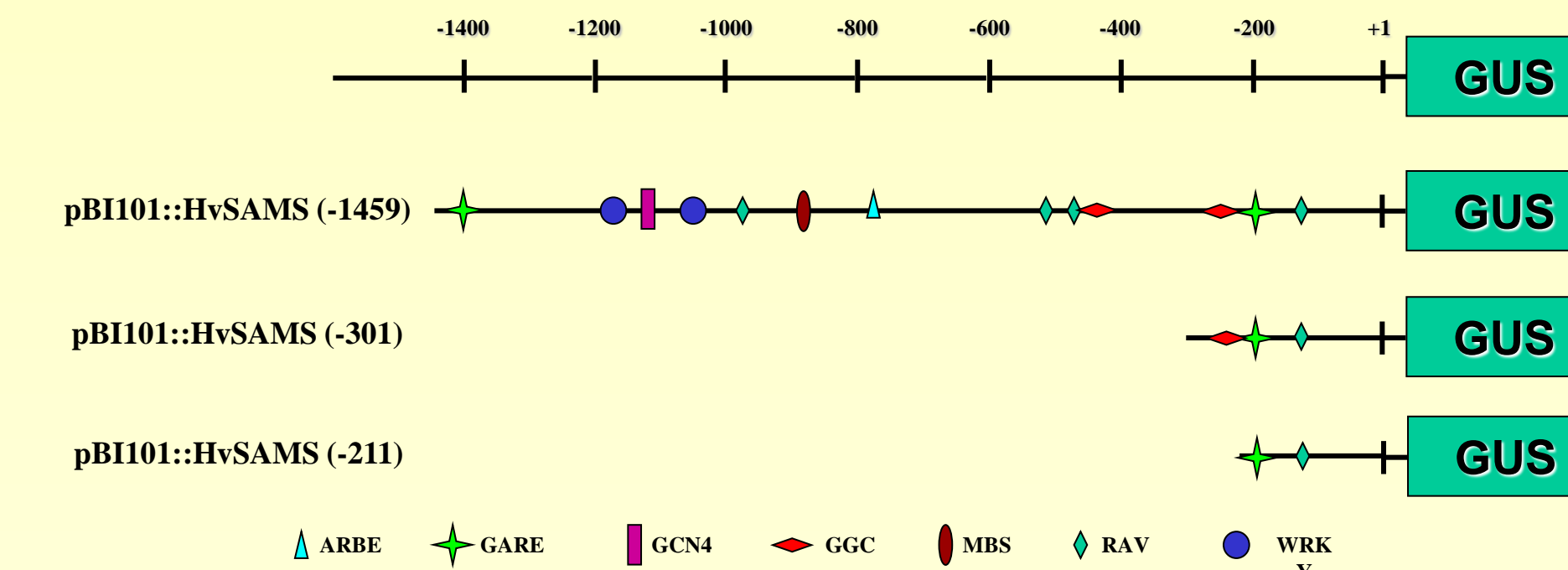
<sup>b</sup> The column was isolated from genome walker.

**Table 2** List of the forward primers used for the RT PCR analysis. The used reverse primers were *HvSAMS 1, 2, 3, and 4* specific primers. Intron 1s were indicated intron regions of the 5' UTR between E1 and E2.

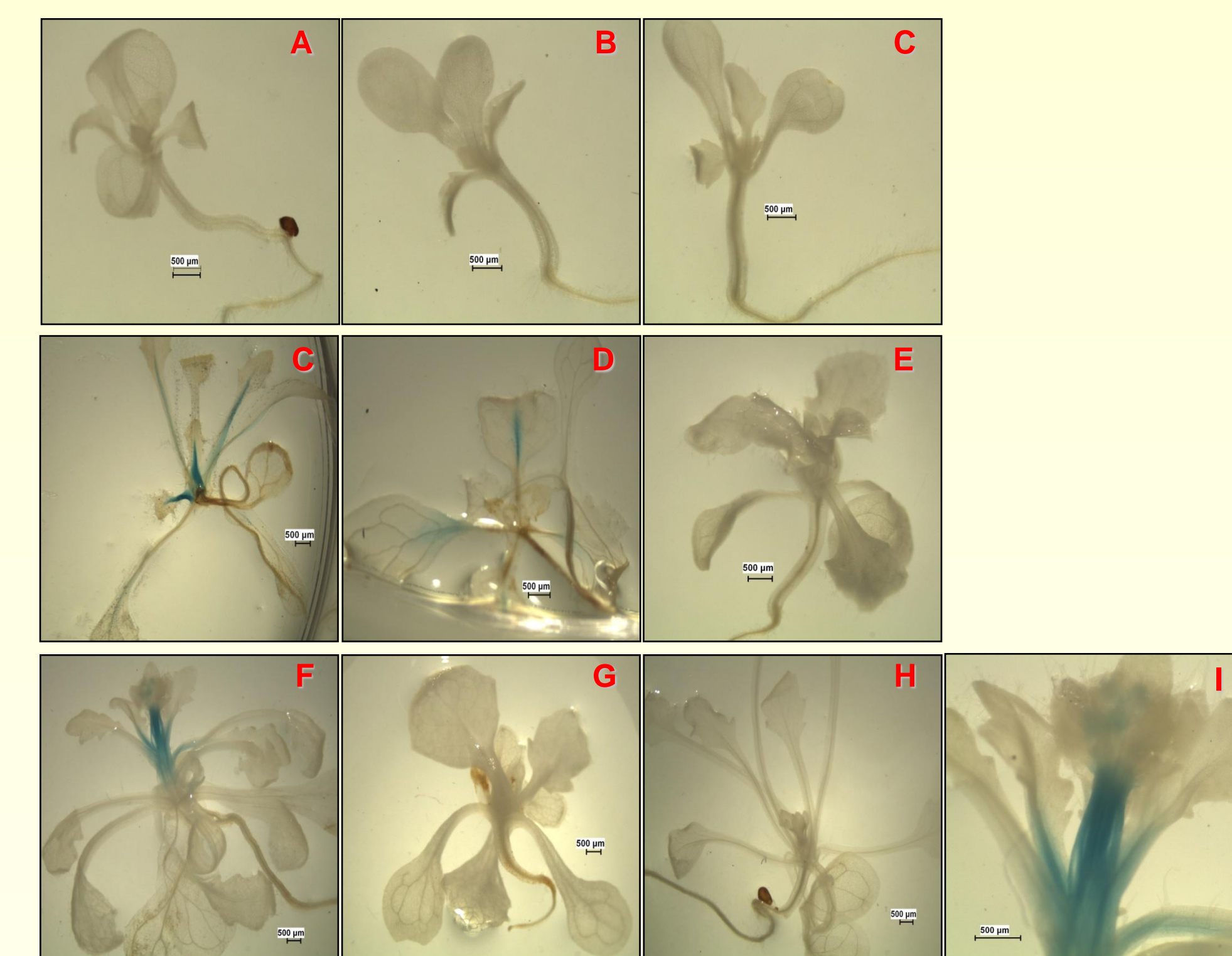
Genes	PCR target	Primer sequence (5' → 3')
<i>HvSAMS 1</i>	Exon1	AGAGCATCACTACCACGAAAG
	Intron1	CTGCTGTATGGCCGGGTAGA
<i>HvSAMS 2</i>	Exon2	TACATTCGACCTTTTCGGTTC
	Intron1	TACATTCGACCTTTTCGGTTC
<i>HvSAMS 3</i>	Exon1	AGAGCATCTCTACCACAAAG
	Intron1	CGTCCTGATCTCATGTTTCGA
<i>HvSAMS 4</i>	Exon2	CTCCTGAACAATAGCATCAGC
	Intron1	GGGTGACCGCTCTCTCTGTTTC
<i>HvSAMS 4</i>	Exon1	AGGCAAAAGATGGCTGAAG
	Intron1	CCTTTCGAGATTGGGTACTG
	Exon2	CCGTGCTGTCTCGAGTGTCTC



**Fig. 2** Transcript accumulation profiles of *HvSAMS 1, 2, 3, AND 4* expressed in plant growth hormones and abiotic stress treatment using RT-PCR analysis. Plant material was 4 week-old leaves of barley treated with a solution of 100 μM each of ABA, GA3, spermidin.



**Fig. 3** Schematic diagram of different *HvSAMS 2* promoter::GUS fusions. The different length promoter fragments were inserted into pBI 121, which was removed CaMV 35S. The putative *cis*-acting elements were represented by symbols.



**Fig. 4** Histochemical localization of GUS activity in transgenic *Arabidopsis* seedlings. *Arabidopsis* Columbia was transformed with transformation vectors. A, C, F, and I, pBI101::HvSAMS (-1459)::GUS fusion vector; B, D, and G, pBI101::HvSAMS (-301)::GUS fusion vector; C, E, and H, pBI101::HvSAMS (-211)::GUS fusion vector. A, B, and C, 1 week-old transgenic plants (T<sub>2</sub>); D, E, and F, 2 week-old transgenic plants (T<sub>2</sub>); F, G, and H, 3 week-old transgenic plants (T<sub>2</sub>); I, 3 times magnified image for F

## Acknowledgement

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