# Integrating ecogeographic, bioclimatic and phylogenetic analyses for the wild relatives of sunflower (Helianthus annuus L.)

1 International Center for Tropical Agriculture (CIAT), Km 17, Recta Cali-Palmira, Cali, Colombia; 2 Biodiversity Boulevard, Vancouver, British Columbia V6T 1Z4, Canada; 3 Department of Agronomy and Plant Genetics, University of Minnesota, 411 Borlaug Hall, 1991 Upper Buford Circle, St. Paul, MN 55108, USA; 4 Centre for Crop Systems Analysis, Wageningen University of Birmingham, Edgbaston, Birmingham B15 2TT, UK; 6 Department of Ecology and Evolutionary Biology, University of Colorado at Boulder, Boulder, CO, USA; 7 USDA-ARS Plant Introduction Research Unit, North Central Regional Plant Introduction Station and Department of Plant Pathology, Iowa State University, Ames, IA 50011, USA; 8 USDA-ARS Northern Crop Science Laboratory, Fargo ND 58102-2765, USA; 9 Department of Biology, Indiana University, Bloomington, IN 1000000, IOWA State University, Ames, IA 50011, USA; 8 USDA-ARS Northern Crop Science Laboratory, Fargo ND 58102-2765, USA; 9 Department of Biology, Indiana University, Bloomington, IN 100000, IOWA State University, Ames, IA 50011, USA; 8 USDA-ARS Northern Crop Science Laboratory, Fargo ND 58102-2765, USA; 9 Department of Biology, Indiana University, Bloomington, IN 100000, 47405, USA \*To whom correspondence should be addressed: <u>kant0063@umn.edu</u>

## Introduction

Helianthus is a genus composed of 63 taxa, including 12 annual and 37 perennial species native to North America, occupying a variety of habitats ranging from open plains to salt marshes (Kane et al., 2013; Seiler & Marek, 2011). Sunflower (*Helianthus annuus* L.) is the most economically important species in the genus, with ~25 million hectares in production worldwide and a substantial private sector breeding effort (Jan & Seiler, 2006; Berglund, 2007; FAOSTAT, 2012). The genepool of sunflower is well defined with most annual taxa hybridizing well while crosses between *H. annuus* and perennial relatives produces mixed results. There are many taxa with adaptive traits such as drought and salinity tolerance. Investigation of the evolutionary history of such taxa is needed in order to prioritize species for conservation and use in sunflower breeding.

## **Objectives**

Prioritize species and geographic areas for further collecting through an examination of 36 *Helianthus* CWR taxa with greatest potential for contribution to sunflower crop improvement

Examine species relationships and niche habitation through comparisons of bioclimatic, ecogeographic, and genetic data to identify potential candidates for use in crop improvement



# **Materials and Methods**

## Results

#### Ecogeography

- 50% of taxa analyzed are in serious need of further collecting
- 28% in moderate need
- 17% were generally well represented in germplasm collections
- There are 5 hotspots of taxon richness (Fig. 1)
- 81% of taxa showed some degree of geographic overlap with wild *H. annuus*

#### Chrystian C. Sosa<sup>1</sup>, Michael Kantar<sup>2,3,\*</sup>, Colin K. Khoury<sup>1,4</sup>, Nora P. Castañeda-Álvarez,<sup>1,5</sup> Nolan Kane<sup>6</sup>, Laura Marek<sup>7</sup>, Gerald Seiler<sup>8</sup>, Harold Achicanoy<sup>1</sup>, Vivian Bernau<sup>1</sup>, and Loren H. Rieseberg<sup>2,9</sup>

- PCA and HCPC (Climate behaviour) Boxplots (90% data)



Fig 1. A) Map showing taxon richness for 36 Helianthus crop wild relatives (CWR). B) Map showing hotspots for proposed collecting of high priority taxa.

## **Bioclimatic analysis**



Fig 2. A) Species occurrence points grouped based on the first three principle components of biophysical and bioclimatic variables. Clusters share homogeneous bioclimatic and biophysical conditions. B) Range variable profiles for soil pH. Boxplot displays the median of 90% of occurrence points for each taxon

- We identified three climatic clusters (Fig. 2A):
- Cluster 1 is defined by dry conditions
- Cluster 2 shows higher values related with precipitation and soil organic carbon
- Cluster 3 represents a combination of soil and temperature variables, with a higher
- than average silt content, a high capacity for cation exchange (CEC), neutral pH, and high soil porosity
- Inclusion of soils variables increased the ability to differentiate species environmental niches
- Niche profiles did display differences between species, e.g. soil pH (Fig. 2B)

#### **Phyloclimatic analysis**

- Mean annual temperature and pH exhibited phylogenetic niche conservatism (Fig. 3)
- The lack of niche conservatism in soil organic carbon and percent sand suggest an important role of soil in the diversification of *Helianthus*

#### Potential candidate taxa

• We identified many different Helianthus taxa that show extreme environmental adaptation and therefore represent potential candidates for use in breeding for climate adaptation (Table 1). These include both new and complementary findings in regard to traits previously reported





Fig. 3. Test of phylogenetic signal with the Blomberg K measure (Blomberg, et al, 2003) using 25 of 36 taxa analyzed with complete genetic and environmental information. K measures phylogenetic signal in traits, where K values below 1 (orange color)indicates low dependence of traits on evolutionary history and K values above 1 indicates trait conservation over evolutionary history. \*indicates K value greater than 1.

Table 1. Selected taxa with history of introgression and potential new traits found using the integrative approach

<u>Taxon</u>	Traits reported	Recommendation	<u>Position in</u> genepool	Environmental Life		Potential extremophile characteristics	
				<u>cluster</u>	<u>history</u>	based on different ecological niche	
		for collection		assignment		relative to H. annuus	
H. anomalus	Fertility restoration	High priority				Tolerance to low precipitation	
					Annual	Tolerance to high pH	
			Secondary	Cluster 3			
	High linoleic		Tertiary	Cluster 1	Perennial	Tolerance to low CEC	
H. atrorubens	acid					Tolerance to high precipitation	
	concentrations	Medium priority					
	in seed					Iderance to low pri	
	(potential)						
	Broomrape		Tertiary	Cluster 3	Perennial		
Н.	resistance;					Tolerance to low temperature	
maximilianii	Cytoplasmic	High priority				Tolerance to erratic temperature	
	male sterility						
	Salt tolerance;	Was assessed to	Secondary	Cluster 1	Annual		
H. paradoxus	fertility	be well				Tolerance to low soil organic carbon	
-	restoration	represented					
H. resinosus	Fertility restoration	Medium priority	Tertiary	Cluster 2	Perennial	Tolerance to high precipitation	
						Tolerance to low CEC	
						Tolerance to low pH	
H. silphioides	High oleic acid	Was assessed to be well	Tertiary	Cluster 2	Perennial		
	concentrations in seed					Tolerance to low CEC	
						Tolerance to low pH	
	(potential)	represented					
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## Conclusions

•The combination of ecological niche and genetic relatedness studies helps identify candidates for adaptive traits

 Examining bioclimatic and biophysical varial geographic parameters add value in identifying further collecting

## Acknowledgements

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#### References

- Berglund, DR (ed.). 2007. Sunflower production. Bull. A-1331 (EB 25 revised). North Dakota State Univ. Ext. Serv., Fargo. • Blomberg S. P., Garland T., Ives A. R.2003 Testing for phylogenetic signal in comparative data: behavioral traits are more labile. Evolution, 57, 717–745..
- FAOSTAT. 2012. Retrieved February, 2014. http://faostat.fao.org. (7 February 2014, date last accessed). Jan, C.-C., & Seiler, G. (2006). Sunflower. In Genetic Resources, Chromosome Engineering, and Crop Improvement (103–165). CRC
- Press. doi:doi:10.1201/9781420005363.ch5 • Kane NC, Burke JM, Marek LF, Seiler GJ, Vear F, Knapp SJ, Vincourt P, and Rieseberg LH. (2013) Sunflower genetic, genomic, and ecological resources. Mol. Ecol. Res. 13: 10-20.
- Ramírez-Villegas, J., Khoury, C., Jarvis, A., Debouck, D. G., & Guarino, L. (2010). A gap analysis methodology for collecting crop genepools: a case study with Phaseolus beans. PloS one, 5(10), e13497.
- Seiler, G., Marek, F. L. (2011). Germplasm resources for increasing the genetic diversity of global cultivated sunflower. Helia, 34(55), 1-20.
- Timme, R. E., Simpson, B. B., & Linder, C. R. (2007). High-resolution phylogeny for Helianthus (Asteraceae) using the 18S-26S
- ribosomal DNA external transcribed spacer. American Journal of Botany, 94(11), 1837-1852. • Warren, D.L., Glor, R.E., Turelli, M., 2008. Environmental niche equivalency versus conservatism: quantitative approaches to niche evolution. Evolution 62 (11), 2868–83.



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