

DNA contents of Texas bluegrass acquired from GRIN, collected in Texas and Oklahoma and interspecific hybrids with Kentucky bluegrass determined by flow cytometry



Jason Goldman



Southern Plains Range Research Station
Woodward, Oklahoma



Texas Bluegrass (*Poa arachnifera*)

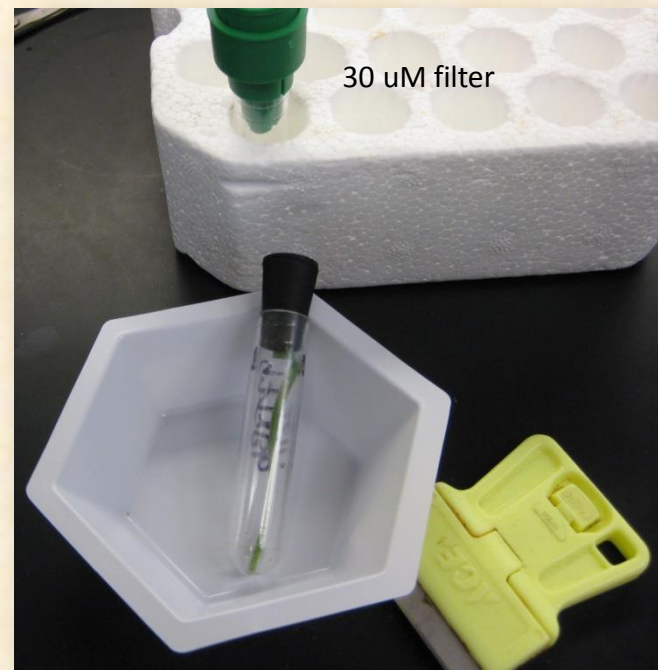
Texas bluegrass is a dioecious native cool season perennial grass that has withstood the region's heat, droughts and overgrazing for centuries. It produces nutritious and palatable forage during the late fall, winter and early spring when most rangeland forages are least preferred by livestock and lowest in nutrients. Interspecific hybrids with Kentucky bluegrass have the potential to produce turf-type material with greater heat tolerance than Kentucky bluegrass. Recently two different reports indicated that Texas bluegrass exists with a range of genome sizes based on flow cytometry measurements. In order to obtain a better idea of the distribution of genome sizes in Texas bluegrass and hybrids, ***the objectives were to use flow cytometry to estimate genome size in:***

- Seeds acquired from GRIN (original submission and increased generation)
- Plants collected in Texas and Oklahoma
- Progeny from controlled crosses between male and female Texas bluegrass plants
- F1 hybrids obtained by controlled crosses between Texas and Kentucky bluegrass
- Advanced generation hybrids

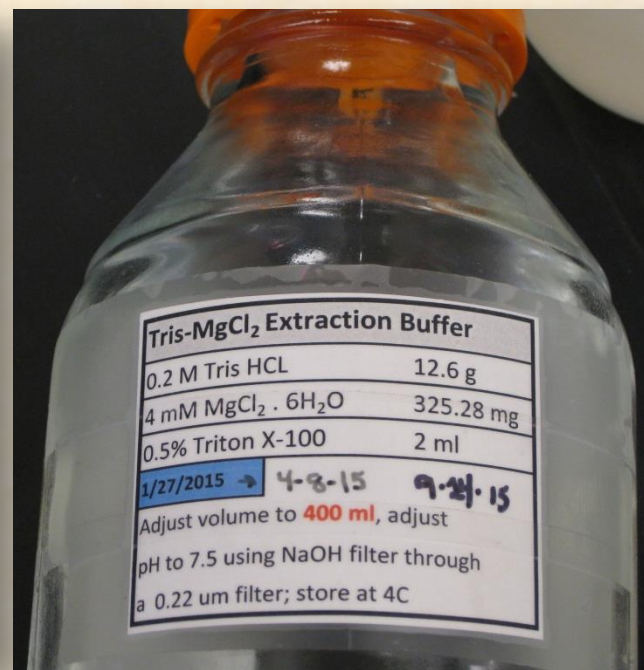




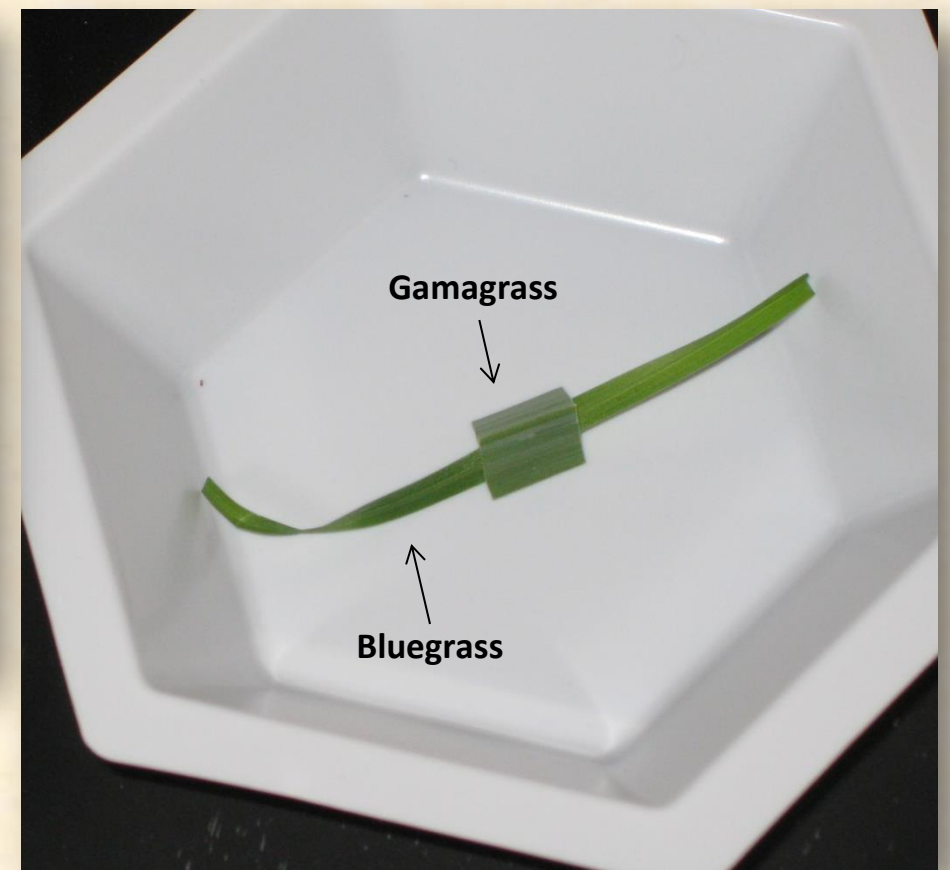
Fresh leaves from plants in the greenhouse or field are placed in small capped tubes and stored in the fridge until processing.



A sharp razor blade housed in a scraper is used to chop leaf tissue in a disposable weigh boat. The extract is filtered through a 30 µm Partec filter.

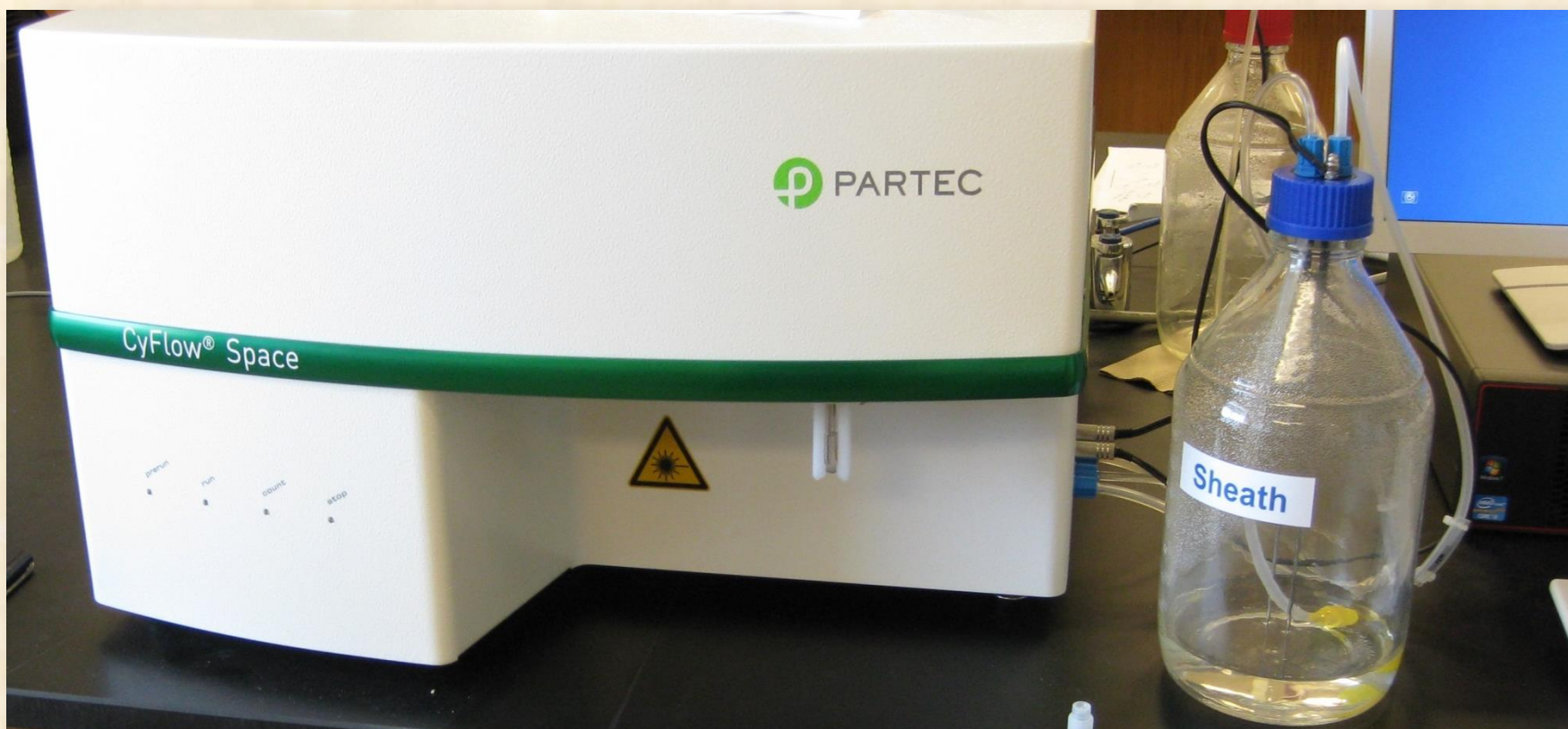


50 µg/mL propidium iodide(PI)
50 µg/mL Rnase
10 mg/mL PVP-40
Added to buffer before use
Buffer kept on ice in the dark



Leaf tissue before the addition of buffer and chopping

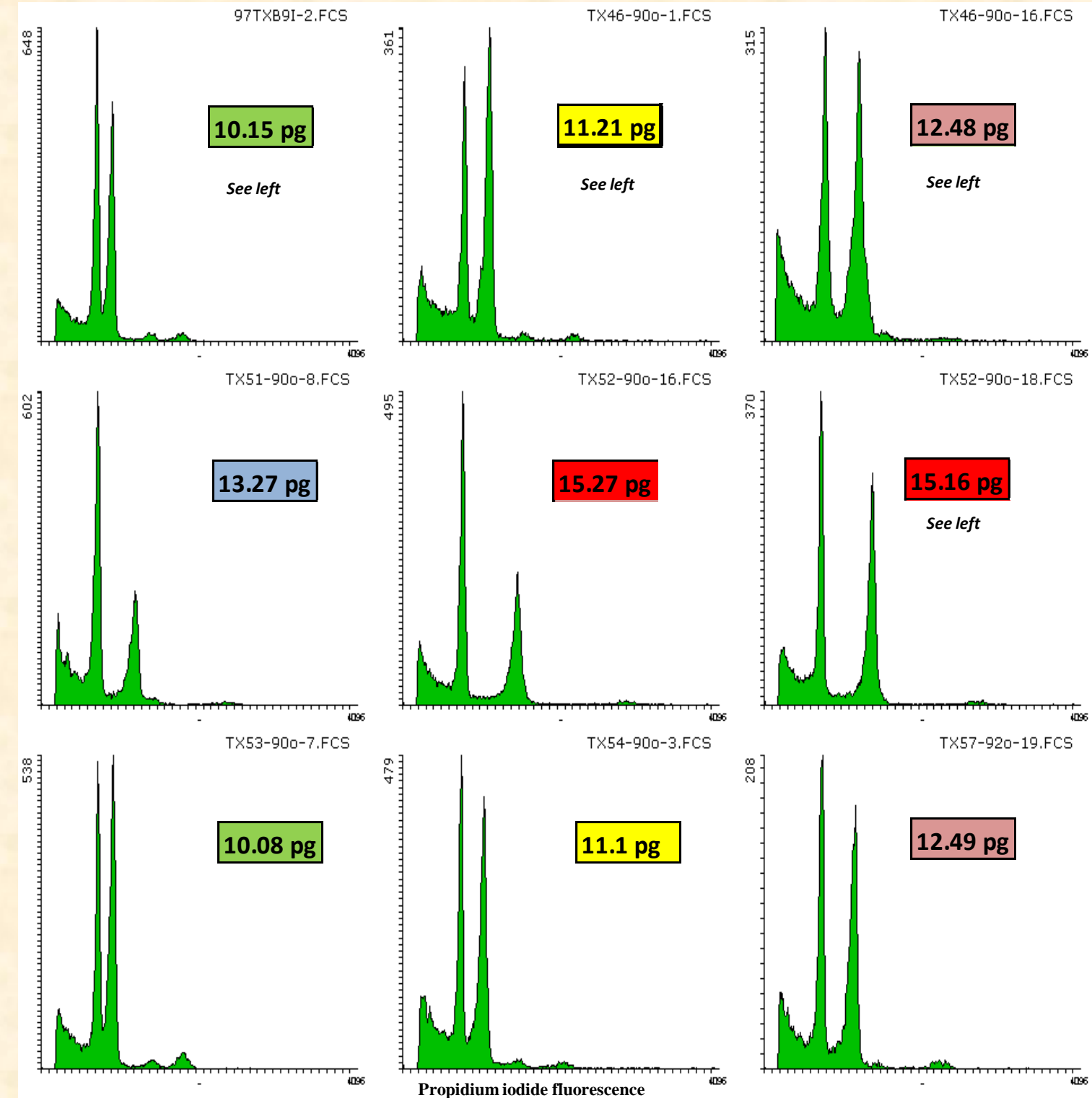
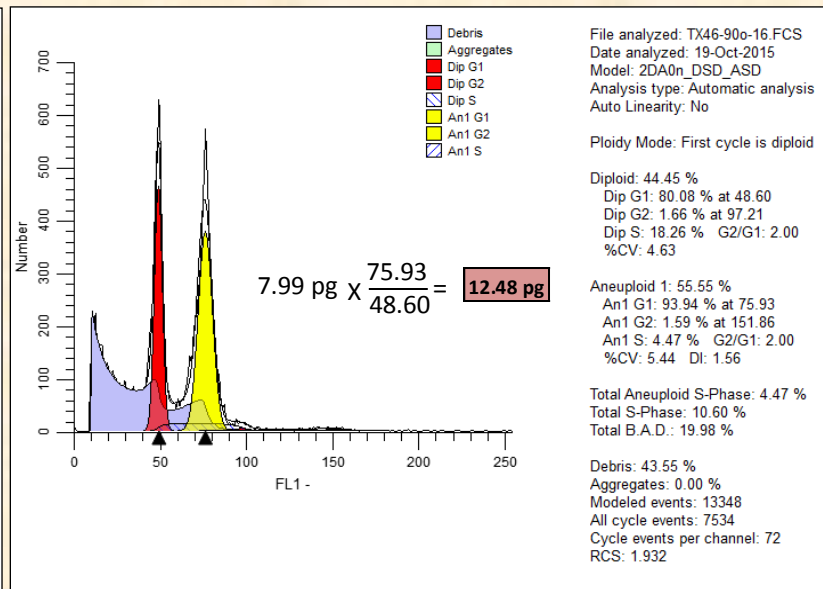
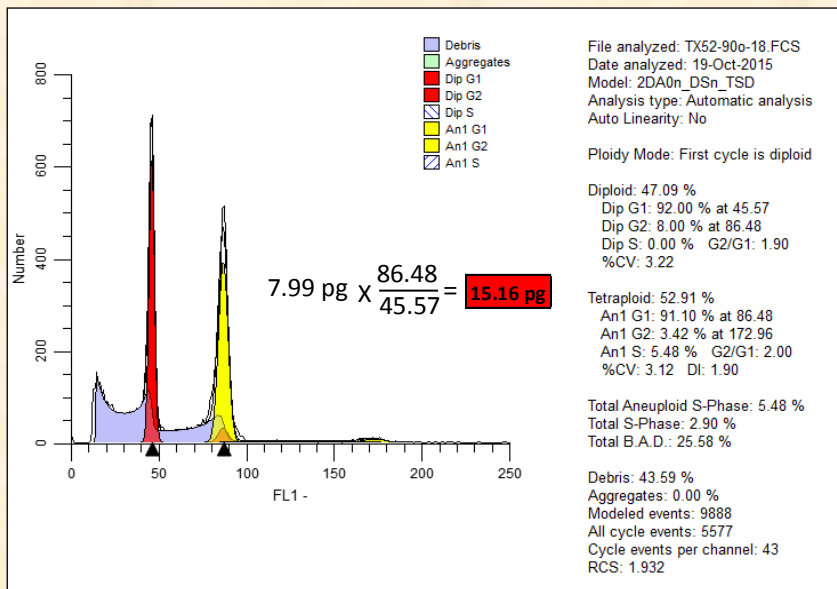
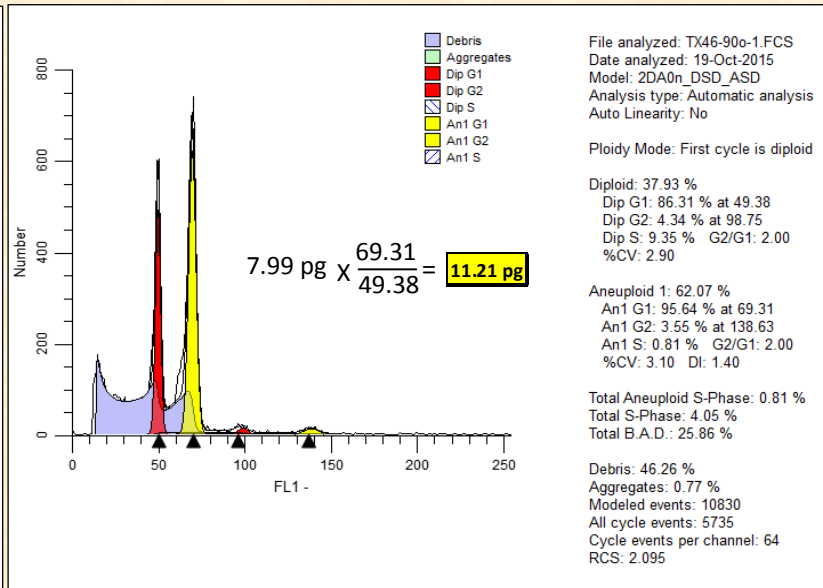
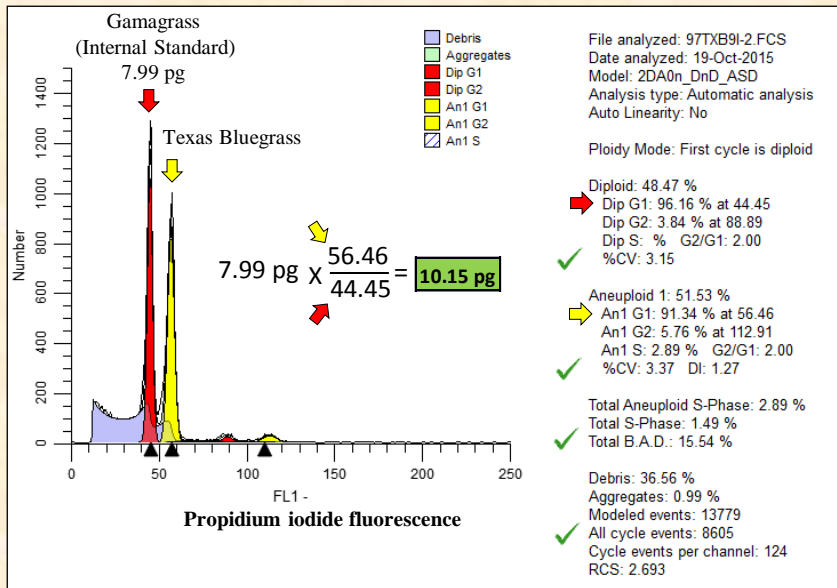
Flow Cytometry



Leaf tissue after the addition of buffer and chopping. The weigh boat is then cupped, shaken, and poured through the green filter.

Samples were processed with a Partec CyFlow Space flow cytometer equipped with a 30 mW 532 nm green laser

Histogram analysis to estimate DNA content



Examples of using Modfit LT 4.1.7 on raw *.fcs files to estimate the mean and **coefficient of variation (CV)** of the peaks of interest and the percentage of **background aggregates and debris (BAD)**. The top left panel shows how peak means are used to calculate the DNA content of Texas bluegrass in picograms (pg) using eastern gamagrass leaf tissue (7.99 pg) as an internal standard.

Examples of histograms produced using eastern gamagrass leaf as an internal standard with Texas bluegrass leaves. The image was created using the raw *.fcs files and the "image collector" function In Flowing Software 2.5.1.

For a precise DNA content estimate the histogram should have:

- ✓ High nuclei counts in peaks of interest
- ✓ Low coefficient of variation (CV) for peaks of interest
- ✓ A low background aggregates and debris (BAD) %

Some further questions to try and answer:

Can multiple DNA contents be detected in Texas bluegrass from sources other than James Read? *Further sampling in progress*

Are there any detectable advantages / disadvantages in the performance of Texas bluegrass individual plants or populations that is related to DNA content?

Are there any advantages/disadvantages related to genome size when selecting female Texas bluegrass plants to be used for creating interspecific hybrids?

Is there a relationship between genome size and any positive or negative agronomic traits when evaluating hybrids derived from crosses between Texas and Kentucky bluegrass for low-input turf?

Acknowledgements:



A portion of this project was supported by a grant from the United States Golf Association, Greens Section Research



I would like to thank members of the Miller lab at Rice University for collecting and supplying Texas bluegrass leaf samples from locations outside northwest Oklahoma



I would also like to thank Vicki Bradley - USDA, ARS Washington State University Regional Plant Introduction Station for supplying the original and increased GRIN seed submissions .