# Broadening of genetic diversity in spring canola *Brassica napus* L. by the use of the C-genome of *Brassica oleracea* L. Xin Wang (MSc.) Supervisor : Habibur Rahman University of Alberta, Edmonton, AB, Canada, T6G 2P5

## Introduction

Rapeseed canola (*Brassica napus* L., AACC, 2n = 38) is one of the important vegetable oilseed crops in the world due to its premium oil quality (Fig 1).

## **Materials and Methods**

Parental material used in this study consisted of; *B. napus* double haploid line A04-73NA *B. oleracea* var. *italica*, broccoli cv. Premium Crop *B. oleracea* var. *capitata*, cabbage cv. Balbro (Fig 2)

## Results

- $F_5$  and  $BC_1S_3$  generation had mean erucic acid 0.18  $\pm$  0.02 SE and 0.27  $\pm$  0.02 SE, respectively which were not significantly different from *B. napus* check parent A04-73NA.
- Mean glucosinolate content in  $F_6$  and  $BC_1S_4$

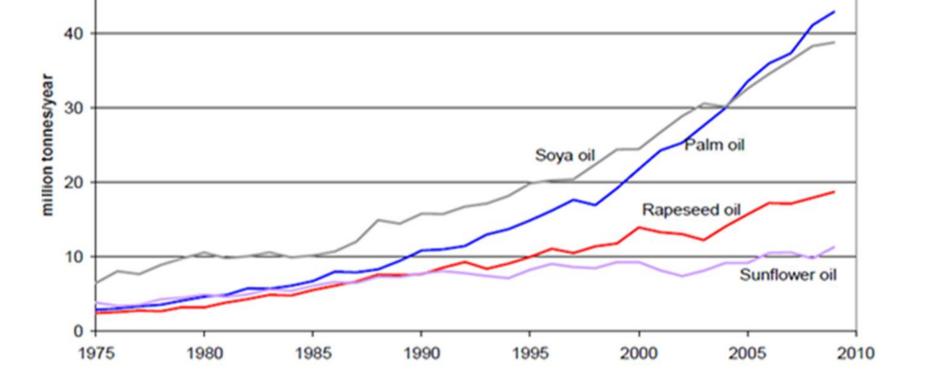


Fig 1. Major vegetable oil production worldwide from 1975 to 2010 (Source: www.fas.usda.gov/psdonline)

Presence of genetic variability is pre-requisite to develop new cultivars with improved yield to meet the demand of ever growing population in the world. However, there has been a decline in genetic diversity of spring canola *B. napus* over last few decades. Therefore, breeding efforts must be taken towards broadening of genetic diversity in spring canola *B. napus* (Cowling 2007, Rahman et al. 2011).

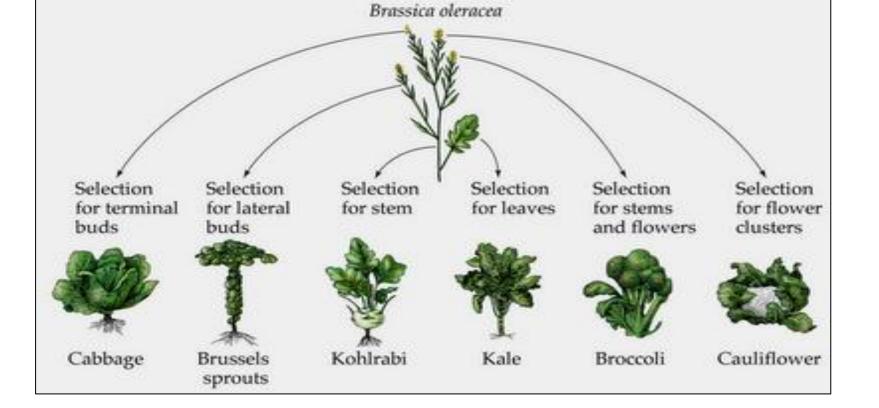


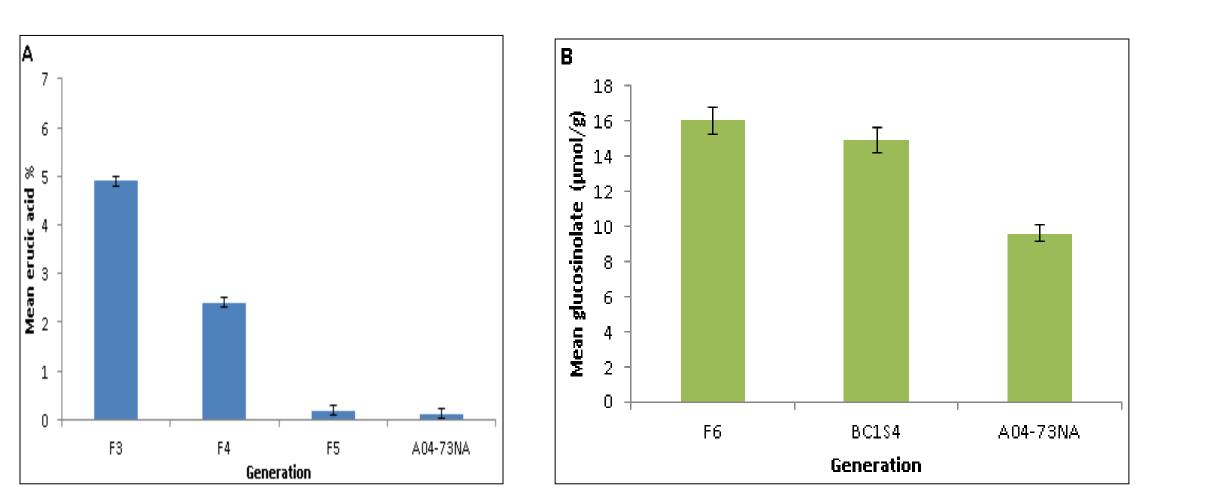
Fig 2. Parental germplasm of *Brassica oleracea* cultivars (Source: www.feralbigten.wordpress.com/2012/02/13/wild-food)

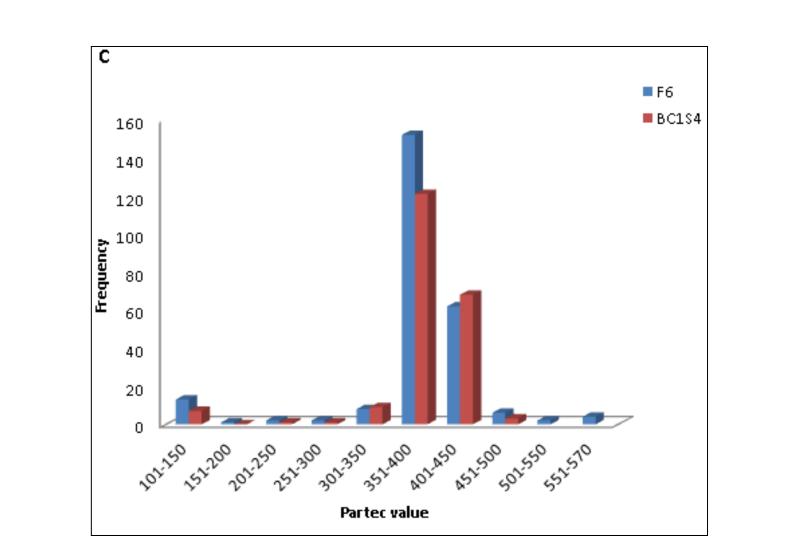
Two types of population were developed according to scheme as shown in Fig 3 which were subjected to evaluation for following traits:

Agronomic traits like days to flowering, silique length (mm), seed number per silique, seed weight (g).

Seed quality traits like fatty acid, oil, protein and glucosinolate contents ( $\mu$  mol g<sup>-1</sup>seed).

- generations were less than 20  $\mu$  mol g<sup>-1</sup>seed.
- Most of inbred lines in  $F_6$  and  $BC_1S_4$  had partec value in the range of *B. napus* check parent A04-73NA.





This can be accomplished by introgressing genetic

diversity from diploid progenitor species *Brassica* rapa L. (AA, 2n = 20), *Brassica oleracea* L. (CC, 2n

= 18) and/or other allied species of the family

### Brassicaceae.

## **Research Objectives**

To assess the feasibility of developing spring canola *B. napus* recombinant inbred lines by crossing *B. oleracea* with *B. napus*.
 To compare the efficiency of filial and backcross breeding method in developing spring canola *B. napus*.



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B. napus × B. oleracea							
A04-73NA) (B. oleracea var. in	(B. oleracea var. italica cv. Premium Crop						
B. oleracea var. c	B. oleracea var. capitata cv. Balbro)						
F1	× B. napus (A04-73NA)						
Self-pollination	$\checkmark$						
F <sub>2</sub>	BC1						
Self-pollination	Self-pollination						
F <sub>3</sub>	BC1S1						
Self-pollination	Self-pollination						
F <sub>4</sub>	BC1S2						
Self-pollination	Self-pollination						
Fn	BC <sub>1</sub> S <sub>n</sub>						
F <sub>2</sub> – derived inbred lines	BC1 – derived inbred lines						

Fig 3. Crossing scheme for interspecific inbred line development



Fig 4. (A) Inheritance of erucic acid in F<sub>2</sub> derived generation;
(B) comparison of glucosinolate in F<sub>6</sub> and BC<sub>1</sub>S<sub>4</sub> generation;
(C) Flow cytometry analysis.
Note: A04-73NA is check *B. napus* parent.

## Conclusion

Several spring canola interspecific *B. napus* inbred lines are developed by crossing *B. napus* with *B. oleracea* which can be used in other breeding programs as well as to develop canola hybrid cultivars.

## References

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