

Quantitative Trait Loci (QTL) Analysis of Freezing Tolerance in Zoysiagrass

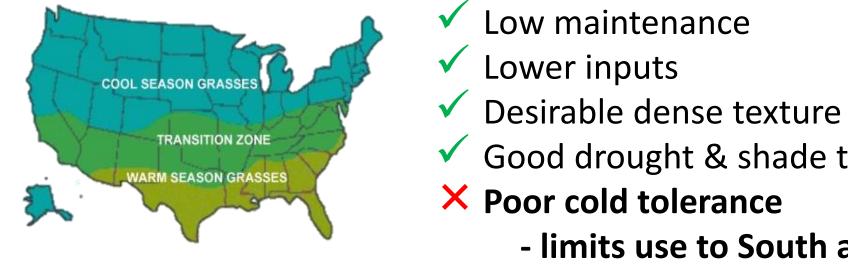


Helen McCamy Pruitt¹, Brian M. Schwartz², Aaron J. Patton³, Consuelo Arellano⁴ and Susana R. Milla-Lewis¹ (1)Crop Science and (4) Statistics, Depts. North Carolina State University, Raleigh, NC (2)Dept. of Crop and Soil Sciences, University of Georgia -Tifton, Tifton, GA (3)Dept. of Horticulture and Landscape Architecture, Purdue University, West Lafayette, IN

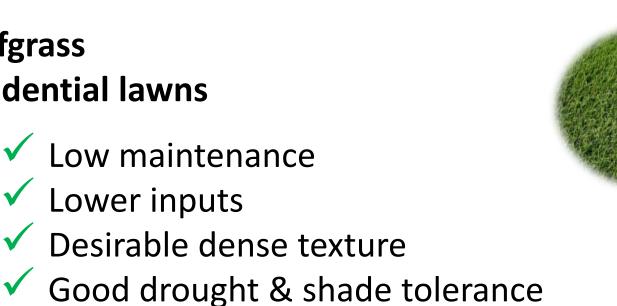
Introduction

Zoysiagrass (Zoysia japonica Steud. and Zoysia matrella (L.) Merr.)

Warm season, perennial turfgrass Used in commercial and residential lawns

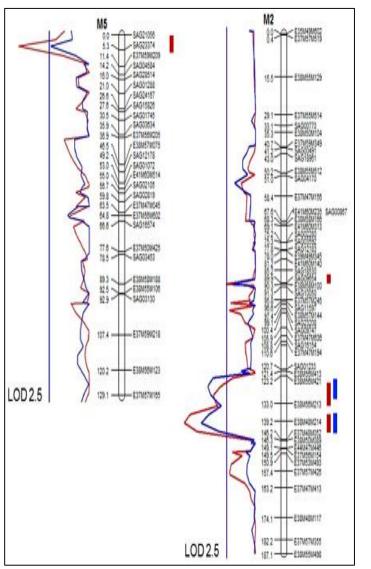


& GENETICS



Molecular Markers Linkage Mapping and QTL Analysis

- Molecular markers are powerful tools for examining genetic variation and constructing linkage maps in plant populations
- Simple sequence repeats (SSRs): Efficient co-dominant PCRbased markers found throughout the genome
- Single nucleotide polymorphisms (SNPs): Abundant molecular markers helpful in creating high density maps



Marker Assisted Selection

Through MAS, the efficiency of selection can be improved by using DNA technology to determine the presence of desirable genes rather than waiting for gene expression. MAS can be used to transfer through crossing and selection specific genes into a desired genetic background.

Research Objectives

1. Phenotype a mapping population of Meyer x Victoria for winter survival

- limits use to South and transition zone

Meyer:

Industry standard for cold tolerance since its release in 1951 **Excellent cold tolerance but poor establishment and coarse** texture



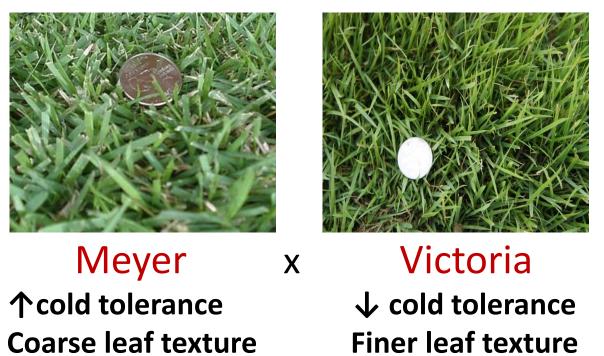
• Genotyping by Sequencing (GBS): Time efficient and cost effective method for rapid SNP discovery

Linkage maps can then be scanned for associations with particular phenotypes in order to identify genomic regions (QTL) controlling the trait of interest.

- 2. Genotype the population using SSR and SNP markers
- 3. Construct a linkage map which will be used to identify QTL controlling winter survival

Materials & Methods

Plant Materials



- 175 progeny
- **Slow establishment**
- Faster establishment
- Pseudo-F₂ mapping population
- Planted in 3 replications in 3x3 ft plots in a randomized complete block design (RCBD) at 3 locations:



• William H. Daniel Turfgrass Research and Diagnostic Center, West Lafayette, IN

Phenotypic Evaluation

- Winter injury, turf quality, percent coverage were evaluated in 2014-2015 and will be evaluated in 2016-2017
- Digital image analysis (FIJI) using pictures taken of each plot periodically from March to October using a lightbox for uniformity of images²
- Winter Injury calculated using the formula: Winter Injury =
 - 100 [(Coverage after Winter Dormancy / Coverage before Winter Dormancy) X 100][°]



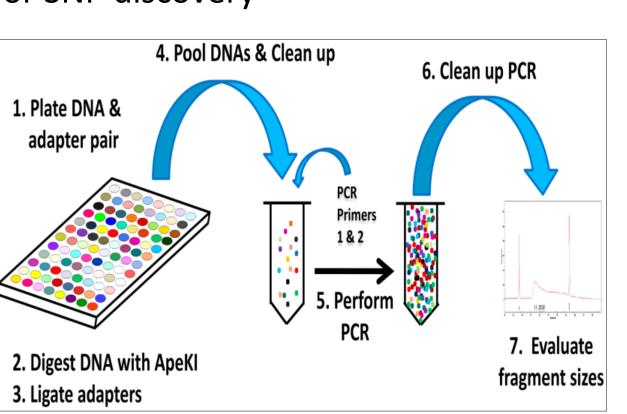
Percent Coverage Evaluation using FIJI

Genotypic Evaluation

1. Plate DNA 8

• **SNPs: Genotyping by Sequencing (GBS)** for SNP discovery ⁶

- Double digest with SphI and MluCl
 - Multiplexed with 96 barcoded adapters
 - adapter pair Four Illumina indices allowed sequencing of
 - 175 progeny + two parents simultaneously
 - Sequence PCR amplified libraries on Illumina HiSeq 200SR platform
 - Unique tags identified in Stacks, compiled, 3. Ligate adapters and used for mapping and SNP calling



Data Analysis

• **SAS**: to evaluate the variability of turf quality, percent cover, and winter survival within and between genotypes⁷

2014 High/Low: 92°F/-21°F • Upper Mountain Research Station,

- Laurel Springs, NC 2014 High/Low: 82°F /-8°F
- University of Georgia Coastal Plain Experiment Station, Tifton, GA 2014 High/Low: 98°F /16°F⁻¹

Genotypic Evaluation

• SSRs

- DNA was extracted from 175 progeny and two parents using CTAB protocol
- The population was genotyped with approximately 125 SSR markers and evaluated on 12% polyacrylamide gel electrophoresis (PAGE) using LICOR sequencers.⁵
- **STACKS:** to call SNPs from 200M+ reads generated by GBS^{*}
- JoinMap: to create a genetic linkage map with approximately 125 SSR markers, potentially over 10,000 SNPs, and the 175 genotypes of the mapping population⁹
- **MapQTL**: to identify QTL for cold tolerance based on genotypic and phenotypic data ¹⁰

Results

Phenotypic Evaluation

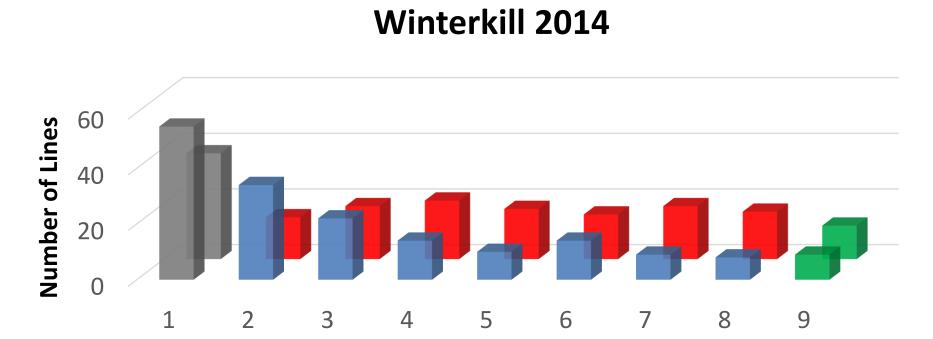
The mapping population showed variation in color, texture, turf quality, aggressiveness, and winter injury in the winter of 2014.



• Winter Survival & Winter Injury

59% winter survival in Laurel Springs, NC **21%** winter survival in West Lafayette, IN

Laurel Sp	rings, NC	West Lafayette, IN
11-TZ-4745	11-TZ-4815	11-TZ-4720
11-TZ-4781	11-TZ-4779	11_T7_//755



Indiana North Carolina

Figure 1: Distribution of winterkill for 175 progeny, Meyer, and Victoria at Laurel Springs, NC and West Lafayette, IN in 2014. Winterkill is a measure of winter injury on a scale of 1 (completely dead) to 9 (no winterkill). Bars in grey and green indicate where Victoria and Meyer, respectively, fell.

Genotypic Evaluation

• SSRs

• 125 primers screened for polymorphisms across the mapping population and will be mapped using JoinMap

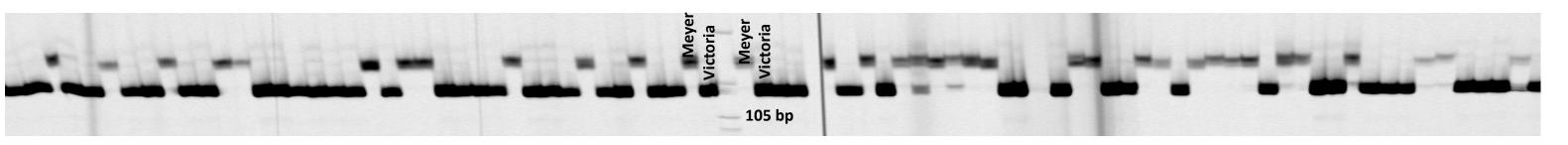
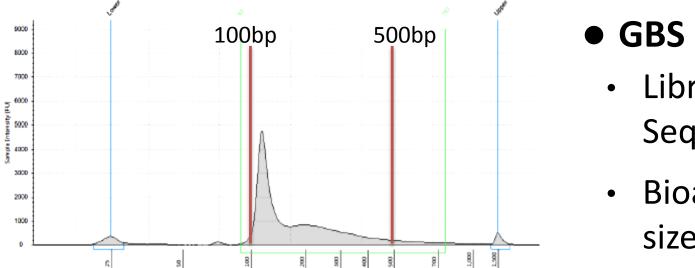


Figure 3: Amplified products generated from SSR b02d15 for selection of individuals from the Meyer x Victoria mapping population. This polyacrylamide gel is an example of screening the population for molecular markers.



- Library was prepped and sent to the NCSU Genomic Sequencing Lab for sequencing
- Bioanalyzer analysis of library shows a concentration of size selected sequenceable fragments at 121bp.

11 12 4/01	II 12 4775	11-12-4/33		
Meyer	11-TZ-4757	11-TZ-4778		
11-TZ-4800	Zenith	11-TZ-4842		
11-TZ-4740	11-TZ-4768	11-TZ-4877		
11-TZ-4758	11-TZ-4840			
11-TZ-4738	11-TZ-4724	Meyer Chinese common		
		Tables 1 & 2: 36 lines suffered no winter		
11-12-4726	11-TZ-4826			
11-TZ-4854	11-TZ-4819	injury in Laurel Springs, NC. Seven lines suffered no winter injury in West Lafayette, IN. Lines that suffered no injury at both		
11-TZ-4794	11-TZ-4727			
11-TZ-4799	11-TZ-4884			
11-TZ-4877	11-TZ-4837			
11-TZ-4784	11-TZ-4753	locations are highlighted in red.		
11-TZ-4851	11-TZ-4789			
11-TZ-4815	11-TZ-4836			
	11-TZ-4800 11-TZ-4740 11-TZ-4758 11-TZ-4758 11-TZ-4738 11-TZ-4720 11-TZ-4843 11-TZ-4854 11-TZ-4854 11-TZ-4799 11-TZ-4799 11-TZ-4877 11-TZ-4851	11-TZ-4800Zenith11-TZ-474011-TZ-476811-TZ-475811-TZ-484011-TZ-475811-TZ-484011-TZ-473811-TZ-472411-TZ-472011-TZ-478211-TZ-472011-TZ-478211-TZ-484311-TZ-489011-TZ-472611-TZ-482611-TZ-479411-TZ-481911-TZ-479911-TZ-488411-TZ-479411-TZ-488411-TZ-485411-TZ-488411-TZ-485111-TZ-475311-TZ-485111-TZ-4789		

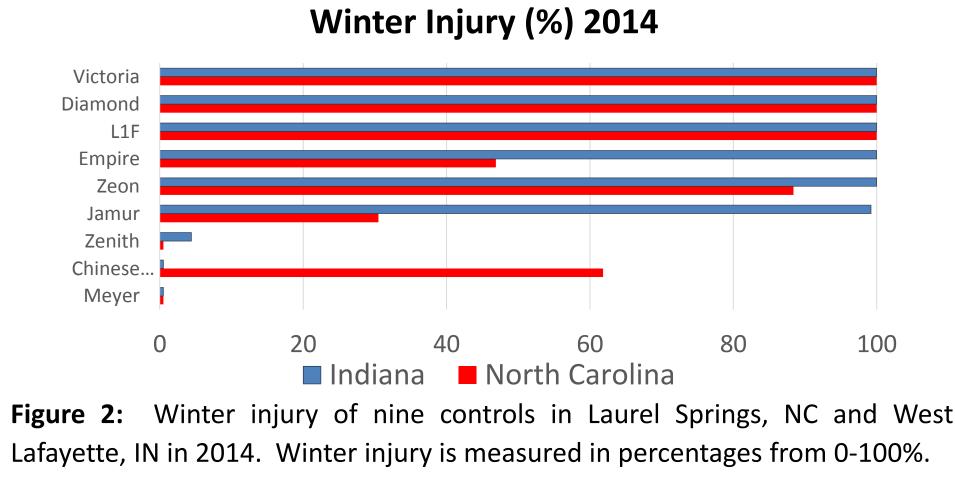


Figure 4: Bioanalyzer results for GBS library

Preliminary Conclusions :: Ongoing Work

• First year data showed wide range of trait expression within the population which is integral to the identification of quantitative trait loci (QTL), so the variability in winter survival seen in these progeny is promising for the identification of markers associated with cold tolerance in zoysiagrass. • Winter injury, turf quality and percent cover data will be collected in 2015 and 2016 Continued genotypic evaluation with SSRs using PAGE, and SNPs using GBS. • The identification and mapping of SNP markers in addition to the SSRs will add depth to the linkage map. • Increased coverage will improve our power to detect QTL for cold tolerance in zoysiagrass.

 National Centers for Environmental Information, National Oceanic and Atmospheric Administration **References:** 2. Richardson, M.D., D.E. Karcher, and K.C. Purcell. 2001. Quantifying Turfgrass Cover using Digital Imaging Analysis. Crop Sci. 41: 1884-1888. B. Patton, A.J. and Z. Reicher. 2007b. Zoysia Winter Hardiness. GCM. 119-123

4. Afanador L, Haley SD, Kelly JD. 1993. Adaption of a "mini-prep" DNA extraction method for RAPD marker analysis in common bean (Phaseolus vulgaris L.). Annual Report of the Bean Improvement Cooperative 36: 10-11. 7. SAS 9.4, SAS Institute, Inc., Cary, NC. 10. MapQTL 6, Kyazma B.V., Netherlands. 5. Wang, D., J. Shi, S.R> Carlson, P.B. Cregan, R.W. Ward. and B.W. Diers. 2003. A Low-Cost, High-Throughput Polyacrylamide Gel Electrophoresis System for Genotyping with Microsatellite DNA Markers. Crop Sci. 43: 1828-1832. 8. JoinMap 4.1, Kyazma B.V., Netherlands 6. Genotyping By Sequencing (GBS). Method Overview. RJ Elshire, JC Glaubitz, Q Sun, JV Harriman. ES Buckler, and SE Mitchell http://www.maizegenetics.net/ 9. Stacks 1.35, Catchen Lab, University of Illinois, 2015

Contact: mhpruitt@ncsu.edu